

Validation of FFQ-based assessment of dietary lignans compared with serum enterolactone in Swedish women

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Abstract

The validity of using FFQ to assess dietary lignans is uncertain. We aimed to validate the use of FFQ for the assessment of dietary intake of lignans compared to the serum biomarker enterolactone, the main product of dietary lignans' metabolism in human subjects. A random sample of women, aged 55–75 years, from the Swedish Mammography Cohort was selected. Information from two FFQ, the FFQ-87 (sixty-seven food items) and the FFQ-97 (ninety-three food items), and blood samples were collected. Dietary intake of lignans (secoisolariciresinol, matairesinol, lariciresinol, pinoresinol, medioresinol and syringaresinol) was assessed by the FFQ. Serum concentrations of enterolactone were analysed by time-resolved fluoroimmunoassay. The correlation coefficient between energy-adjusted lignan intake and serum enterolactone was estimated in crude and multivariable-adjusted models, taking into account the factors potentially influencing the serum enterolactone. Among the 135 participants aged 55–75 years, with a mean BMI of 26.7 kg/m², the average energy-adjusted intake of total lignans was 1616 (SD 424) and 1516 (SD 409) µg/d according to the FFQ-87 (forty-five food items containing lignans) and the FFQ-97 (sixty-five food items containing lignans), respectively. The mean concentration of serum enterolactone was 23.2 (SD 15.4) nmol/l. The adjusted Pearson's correlation between dietary intake of lignans assessed by the FFQ-97 and serum enterolactone was statistically significant (r 0.22, $P=0.01$). No significant correlation was observed for the FFQ-87 (r 0.09, $P=0.30$). The present study indicates that the FFQ-97 might be better than the FFQ-87 for assessing dietary intake of lignans, although the correlation was low.

Key words: Plant oestrogens: Validity: Biological markers: Diet: Lignans

Phyto-oestrogens are naturally occurring polyphenolic plant compounds with hormone-like activity^(1,2). Lignans, present in, for example, wholegrain products, berries, fruits, vegetables, flaxseed and sesame seed, are the most abundant phyto-oestrogens in Western diets⁽³⁾. After consumption, lignans are converted by the gut microflora into the so-called enterolignans, enterolactone and its immediate precursor enterodiol^(2,4,5). Until now, six plant lignans have been identified as precursors of enterolactone, including secoisolariciresinol (SEC), matairesinol (MAT), lariciresinol (LAR), pinoresinol (PIN), medioresinol (MED) and, to a minimal extent,

syringaresinol (SYR)⁽³⁾. Some studies have indicated that dietary phyto-oestrogens protect against CVD^(6,7), breast cancer^(8–10), prostate cancer^(11–13) and menopause-related symptoms^(14–16), but not all^(17,18). A possible reason for this inconsistency is the measurement error of phyto-oestrogen intake based on FFQ^(19–24). For instance, the long-term intake estimation of lignans based on FFQ might be subject to within-person random error due to the day-to-day fluctuation in dietary intake. In addition, systematic between-person errors might also have been produced due to the omission of lignan-containing food items in a standardised FFQ or the

Abbreviations: LAR, lariciresinol; MAT, matairesinol; MED, medioresinol; PIN, pinoresinol; SEC, secoisolariciresinol; SYR, syringaresinol.

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lack of a complete lignan composition database. Due to these measurement errors, the estimated level of lignan intake among study subjects might be inaccurate and therefore dilute the true association with disease⁽²⁵⁾. Four validation studies of dietary phyto-oestrogens have compared FFQ with relevant biomarkers^(11,26–28). One Finnish study and one Swedish study of men indicated significant correlations (both *r* 0.19) when comparing the FFQ-based estimate with the serum concentration of enterolactone, the biomarker of lignan intake^(11,26). However, two English studies did not observe a correlation between dietary intake of lignan and serum concentration of enterolactone^(27,28). The purpose of the present study was to assess the validity of two versions of FFQ, the FFQ-87 with a shorter and the FFQ-97 with a longer list of food items containing lignans, in the measurement of dietary intake of lignans, as compared to serum concentrations of enterolactone, among Swedish women.

Method

Study subjects and study design

The present validation study included women randomly drawn from the Swedish Mammography Cohort. The Swedish Mammography Cohort was established between 1987 and 1990. All 90 303 women born between 1914 and 1948 and residing in the Västmanland and Uppsala counties in central Sweden were invited by mail to participate in a population-based mammography-screening programme. Enclosed with this invitation was the FFQ-87 that elicited information on diet, weight, height and education. The participation rate was 74%. In 1997, a more comprehensive dietary questionnaire, the FFQ-97, was sent to the 56 030 Swedish Mammography Cohort members who were residing in the study area, and 70% of them returned the completed questionnaire. During 2003–4, 140 women aged 55–75 years were randomly selected from the cohort for the present validation study. None of them had used antibiotics in the past year. This exclusion was necessary as antibiotics are known to influence phyto-oestrogens' metabolism⁽²⁹⁾. All 140 participants answered the FFQ-87 and the FFQ-97 in 2003–2004, and fasting blood samples were collected within 3 months of completing these questionnaires. The information on history of gastrointestinal disease was obtained from the National Patient Register, while the diabetes data was collected from the combination of the National Patient Register, the National Diabetes Register and self-reported questionnaires. The study was approved by the Regional Ethics Committee at the Karolinska Institutet in Stockholm, Sweden.

Assessment of dietary phyto-oestrogen intake using FFQ

The FFQ-87 and the FFQ-97 included sixty-seven and ninety-three food items, respectively. In the FFQ-87, participants were asked to report their average frequency of consumption for each type of food or beverage using eight predefined frequency categories: 'never/seldom', 'one to three times/month', 'one time/week', 'two to three times/week', 'four to six times/

Table 1. Dietary lignan intake among 135 Swedish women (Mean values and standard deviations; medians and ranges)

Dietary intake (µg/d)†	No. of participants	FFQ-87*				FFQ-97*				P (FFQ-87 v. FFQ-97)
		Mean	SD	Median	Range	Mean	SD	Median	Range	
Total lignans	135‡	1616	424	1552	696–2863	1516	409	1497	597–3451	0.002
Matairesinol		29	13	27	5–74	41	17	39	10–105	< 0.0001
Secoisolariciresinol		112	38	105	44–237	157	44	151	71–338	< 0.0001
Lariciresinol		484	128	477	208–846	495	170	480	134–1127	0.43
Pinoresinol		676	227	648	177–1473	510	180	481	138–1227	< 0.0001
Syringaresinol		288	83	290	75–475	278	103	261	128–735	0.29
Medioresinol		24	7	25	9–39	35	14	32	13–97	< 0.0001
Total lignans by BMI (kg/m ²)§										
< 25	54	1542	399	1529	696–2477	1464	368	1509	807–2323	0.14
25.0–29.9	50	1664	373	1626	1031–2810	1522	303	1482	801–2251	0.007
≥ 30.0	29	1686	543	1507	849–2863	1634	589	1638	597–3451	0.52
Total converted lignans	135‡	992	257	951	446–1762	958	279	939	306–2092	0.001

* FFQ-87: FFQ, 1987 version (forty-five food items containing lignans); FFQ-97: FFQ, 1997 version (sixty-five food items containing lignans).

† Nutrient intakes are energy-adjusted via the residual method.

‡ Five outliers (95% CI) were excluded.

§ Two subjects had missing BMI values.

|| Expected amount of dietary lignans was converted to enterolactone in the intestine using conversion factors: matairesinol = 0.62, secoisolariciresinol = 0.72, lariciresinol = 1.01, pinoresinol = 0.55, syringaresinol = 0.44, pinoresinol = 0.55, syringaresinol = 0.04 and medioresinol = 0.8.

week', 'one time/d', 'two to three times/d' or 'four times/d'. In the FFQ-97, close-ended questions with similar response categories were set for most food items, but open-ended questions (open answers, not pre-specified categories) were designed for some commonly consumed foods including bread, milk, cheese, soft drinks, beer, coffee, tea and sugar. Energy content was obtained from the Swedish National Food Administration database⁽³⁰⁾. Total lignan intake was estimated using published content values of the six most prevalent dietary precursors of enterolactone: SEC, MAT, LAR, PIN, MED and SYR^(2,31–37). Of the sixty-seven and ninety-three food items listed in the FFQ-87 and the FFQ-97, forty-five (69.2%) and sixty-five (69.9%) items were assigned lignan values, respectively. The remaining had no values assigned because the lignan content was assumed to be negligible. Nutrient intake was computed by multiplying the frequency of food items by the nutrient content of the age-specific servings. The estimations of total intake of lignans were adjusted for total energy intake, using the residual method⁽³⁸⁾. As the activity of the gut microflora influences the metabolism of dietary lignans to enterolactone^(4,5,39), we also used a formula based on experimental results to calculate the expected amount of mammalian lignans, which had been converted from dietary lignans⁽⁴⁰⁾:

$$\begin{aligned} \text{Estimate of enterolactone} = & 0.62 \times \text{MAT} + 0.72 \times \text{SEC} + 1.01 \\ & \times \text{LAR} + 0.55 \times \text{PIN} + 0.04 \times \text{SYR} \\ & + 0.8 \times \text{MED}. \end{aligned}$$

Analysis of serum enterolactone

Blood samples were processed and separated for sera that were stored at -80°C until analysis. Samples were shipped frozen to the Folkhälsan Institute for Preventive Medicine, Nutrition and Cancer (Helsinki, Finland), where they were thawed and subjected to overnight enzymatic hydrolysis and diethyl ether extraction. Sample extracts were then diluted in assay buffer, with europium label, internal standards and subsequently analysed by time-resolved fluoroimmunoassay according to previously reported protocols for assessment of enterolactone^(41–43). The intra- and inter-assay CV % of the time-resolved fluoroimmunoassay method was low (3.3–6.0 and 6.9–9.9% for enterolactone, depending upon the serum concentrations)^(41–43). Serum isoflavone genistein was analysed using the same method, but only forty of the total 140 women had a detectable serum concentration within the range 0–48 (median 3) nmol/l, indicating low consumption of the isoflavone genistein in the Swedish diet. Therefore, genistein was not included in the final analysis.

Statistical analysis

Lignan values were not normally distributed; therefore, the log-transformed values of lignan intake and serum enterolactone were used in the Pearson's correlation analyses. A total of five observations outside the 95% CI of the corresponding values were excluded, leaving 135 participants for final analysis. The following variables were considered as potential

Table 2. Serum concentration of enterolactone by characteristics of study subjects (Mean values and standard deviations; medians and ranges)

Serum concentration (nmol/l)	No. of participants	Mean	SD	Median	Range
Total population	135*	23.2	15.4	20.8	0.7–68.4
Age group (years)					
55–62	40	21.6	14.2	19.5	0.7–68.4
63–69	58	22.6	15.4	20.3	1.4–62.6
≥ 70	37	25.8	16.7	25.2	1.5–60.1
<i>P</i> †		0.46			
BMI (kg/m ²)‡					
< 25	54	23.1	13.1	21.6	1.5–58.2
25.0–29.9	50	25.8	17.9	22.8	0.7–68.4
≥ 30.0	29	18.4	13.4	17.4	1.4–56.5
<i>P</i> †		0.13			
Constipation					
Yes	14	19.6	11.9	21.7	5.3–48.5
No	121	23.6	15.8	20.8	0.7–68.4
<i>P</i> §		0.36			
Diabetes					
Yes	7	17.6	17.1	9.1	3.7–47.9
No	128	23.7	15.3	21.3	0.7–68.4
<i>P</i> §		0.32			
Gastrointestinal disease					
Yes	14	20.9	10.9	22.7	2.3–39.4
No	121	23.5	15.9	20.5	0.7–68.4
<i>P</i> §		0.55			

* Five outliers (95% CI) were excluded.

† ANOVA was used to compare the mean values.

‡ Two subjects had missing BMI values.

§ The *t* test was used to compare the mean values.

Table 3. Food sources for major contribution of dietary intake of lignans among 135 Swedish women

Food sources	Syringaresinol (%)	Medioresinol (%)	Pinoresinol (%)	Lariciresinol (%)	Matairesinol (%)	Secoisolariciresinol (%)	Total lignans (%)
Wholemeal bread	46	51	22	13	26	6	27
Crisp bread	25	18	26	14	56	12	25
White bread	14	16	3	2	3	0.9	6
Vegetables	0.2	2	28	41	1	9	14
Broccoli	–	–	6	16	0.6	3	4
Cabbage	–	–	17	8	–	0.6	4
Mixed vegetables	–	–	2	7	0.08	0.4	1
Fruits/berries	8	8	10	16	3	23	11
Orange/citrus fruits	7	4	3	7	–	4	4
Apple	–	–	1	6	0.4	1	2
Berries	–	–	0.4	0.4	1	9	2
Coffee	–	–	0.2	1	–	18	3
Flakes	2	2	6	3	2	2	3
Carrots	–	–	0.9	3	0.00	7	2
Wine	–	–	0.02	0.2	3	6	2

confounders, as they might influence the metabolism of dietary lignans: age (categorised into three groups: 55–61, 62–69 and >69 years), BMI (≤ 24.9 , 25–29.9 and ≥ 30 kg/m²), constipation (yes or no), gastrointestinal disease history (yes or no) and diabetes (yes or no). The analyses were implemented in both crude and multivariable models. The basic model included adjustment for age only, while the full multivariable model included all variables listed earlier. The partial Pearson's correlation was used to calculate the adjusted correlation coefficients in the full model. Test of linear trend across BMI categories was conducted by assigning the median of BMI in each BMI category and then treating these values as a continuous variable in the model. As previous studies have indicated BMI as an important determinant of the serum enterolactone concentration⁽⁴⁴⁾, additional analyses stratified by BMI subgroups were also conducted. ANOVA and *t* tests were used to compare the mean values of serum concentration of enterolactone in the subgroups when appropriate. All statistical analyses were performed using SAS 9.0 (Statistical Analysis System, version 9.0; SAS Institute).

Results

Dietary intake of lignans and serum enterolactone

Among the 135 study participants aged 55–75 years, with a mean BMI of 26.7 kg/m², the average energy-adjusted dietary intake of total lignans was 1616 (SD 424) µg/d according

to the FFQ-87 and 1516 (SD 409) µg/d for the FFQ-97 (Table 1). The estimates of dietary lignan intake were different for the two FFQ ($t = 3.2$, $P = 0.002$). The average intake of lignans using the FFQ-97 in obese subjects (BMI ≥ 30 kg/m²) was 1634 (SD 589) µg/d and 1464 (SD 368) µg/d in normal-weight (BMI < 24.9 kg/m²) subjects ($t = -1.42$, $P = 0.16$). The mean concentration of serum enterolactone was 23.2 (SD 15.4) nmol/l (Table 2). No statistically significant difference of serum enterolactone was observed among subgroups of BMI, age groups, constipation, diabetes and gastrointestinal disease history. Approximately 60% of the daily intake of total lignans was derived from bread, while fruit and vegetable intake accounted for 25% (Table 3). Lignan LAR (32.4%) and PIN (33.3%) were the main contributors of total lignan intake (data not shown).

Correlation between dietary lignans and serum enterolactone

Pearson's correlation coefficients between lignan intake and serum concentration of enterolactone are presented in Table 4. Dietary lignans assessed by the FFQ-97 were statistically significantly correlated with serum enterolactone in the crude and multivariable-adjusted models ($r = 0.16$, $P = 0.06$; $r = 0.22$, $P = 0.01$, respectively), as well as when converted lignan values were used ($r = 0.17$, $P = 0.04$; $r = 0.24$, $P = 0.006$, respectively) (Table 4). However, no such correlation was found for the FFQ-87 with lignan intake ($r = 0.09$, $P = 0.30$), nor with

Table 4. Pearson's correlation between energy-adjust dietary intake of lignans based on the FFQ and serum concentration of enterolactone*

Intake	<i>n</i>	Serum v. FFQ-87†				Serum v. FFQ-97†				FFQ-87 v. FFQ-97	
		Crude	<i>P</i>	Adjusted‡	<i>P</i>	Crude	<i>P</i>	Adjusted‡	<i>P</i>	Crude	<i>P</i>
Total lignans	135§	0.06	0.47	0.09	0.30	0.16	0.06	0.22	0.01	0.59	< 0.0001
Converted lignans	135§	0.08	0.33	0.12	0.19	0.17	0.04	0.24	0.006	0.62	< 0.0001

* All dietary intakes of lignans and serum enterolactone were log-transformed.

† FFQ-87: FFQ, 1987 version (forty-five food items containing lignans); FFQ-97: FFQ, 1997 version (sixty-five food items containing lignans).

‡ Adjusted for age, BMI (<25, 25–29.9, ≥ 30) kg/m², constipation (yes/no), gastrointestinal disease history (yes/no) and diabetes (yes/no).

§ Five outliers (95% CI) were excluded.

|| Expected amount of dietary lignans was converted to enterolactone in the intestine using conversion factors: matairesinol = 0.62, secoisolariciresinol = 0.72, lariciresinol = 1.01, pinoresinol = 0.55, syringaresinol = 0.44, pinoresinol = 0.55, syringaresinol = 0.04 and medioresinol = 0.8.

converted lignans (r 0.12, $P=0.19$). The correlation between the two FFQ was statistically significant (r 0.59, $P<0.0001$).

Pearson's correlations between intake of total lignans and of converted lignans and serum enterolactone in BMI subgroups are shown in Table 5. The highest correlation was observed in obese women ($BMI \geq 30 \text{ kg/m}^2$). A borderline positive trend between increasing BMI and increasing correlation coefficients for lignans was indicated, but due to the limited sample size, chance errors cannot be ruled out.

Discussion

The present study shows that the validity of FFQ-based estimates of dietary lignan intake depends on the number of food items containing lignans in the FFQ, and potentially the BMI of the study subjects.

Limitations of the present study include the small sample size, although it was not smaller than that of other phyto-oestrogen validation studies^(11,28,45). A limitation of the serum enterolactone assessment was that only a single fasting blood sample was used. The high intra-individual variation with poor precision when using a single measurement might lead to misclassification of the enterolactone exposure⁽⁴⁶⁾. However, the concentration of serum enterolactone in the present study was in line with the concentrations observed in other Swedish studies^(47,48). Among strengths were the adjustments for confounders, including diabetes and gastrointestinal diseases, that might interact with phyto-oestrogens' metabolism through enzymatic modification and gut microflora⁽⁴⁹⁻⁵¹⁾. Furthermore, none of the participants had used antibiotics during the previous 12 months, which might otherwise be a matter of concern⁽²⁹⁾. The use of antibiotics is known to reduce the amount of gut bacteria and subsequently serum concentrations of enterolactone⁽⁵²⁾. Additionally, the use of fasting blood samples was an advantage, as non-fasting samples might have lower reliability regarding enterolactone measurements^(46,47,53). Another strength was the inclusion of four newly identified enterolactone precursors (LAR, PIN, SYR and MED) in the calculation of total dietary lignan intake, while most of the previous studies included only SEC and MAT for such calculations⁽²⁶⁻²⁸⁾.

Few studies^(11,26-28) have investigated the correlation between lignan intake estimates and serum enterolactone as a biomarker (Table 6). In a study of 140 women in England, the correlation between dietary intake measured by FFQ and serum enterolactone was low (r 0.10, $P=0.20$)⁽²⁷⁾. However, a Finnish study showed better validity of 24 h recalls (r 0.19, $P<0.0001$) in a large study population (n 1784), but they assessed only the precursors SEC and MAT to estimate lignan intake⁽²⁶⁾. The only previous study that estimated total lignan intake based on all six precursors of plant lignans was performed in Swedish men, and showed a correlation of 0.19 ($P=0.09$)⁽¹¹⁾. The correlation observed in the present study of the FFQ-97 (r 0.22, $P=0.01$) was similar.

The reasons for the low correlation between lignan intake and serum enterolactone might be diverse. FFQ might overestimate the consumption of a number of food groups, particularly lignan intake from fruits and vegetables⁽³⁸⁾, and

Table 5. Pearson correlations between energy-adjusted dietary intake of lignans based on the FFQ and serum concentration of enterolactone stratified by BMI*

BMI (kg/m ²)	Serum v. FFQ-87†			Serum v. FFQ-87_converted‡			Serum v. FFQ-97†			Serum v. FFQ-97_converted‡		
	n	Crude	P	Crude	P	Adjusted§	Crude	P	Adjusted§	Crude	P	Adjusted§
< 25	54	0.007	0.96	0.04	0.76	0.03	0.13	0.35	0.14	0.13	0.35	0.14
25.0-29.9	50	0.11	0.47	0.11	0.44	0.12	0.08	0.57	0.14	0.08	0.60	0.19
≥ 30.0	29	0.13	0.50	0.15	0.45	0.27	0.36	0.06	0.38	0.37	0.05	0.38
P for trend¶			0.15		0.15	0.13		0.09			0.12	

* All dietary intakes of lignans and serum enterolactone were log-transformed.

† FFQ-87: FFQ, 1987 version (forty-five food items containing lignans); FFQ-97: FFQ, 1997 version (sixty-five food items containing lignans).

‡ Expected amount of dietary lignans could be converted to enterolactone in the intestine.

§ Adjusted for age, constipation (yes/no), gastrointestinal disease history (yes/no) and diabetes (yes/no), when applicable.

|| Two subjects had missing BMI values.

¶ Test for trend across BMI categories was conducted by assigning the median of BMI in each BMI category and then treating these values as a continuous variable.

Table 6. Summary of papers investigating correlation between dietary intake of lignans and serum concentration of enterolactone

Authors	Study population	Dietary measurement	Estimate of lignan intake	Serum enterolactone analysis	Statistical analysis	Correlation coefficient	
						r	P
Lin <i>et al.</i> (present study)	Swedish women (n 135)	FFQ-97 (ninety-three food items)	Matairesinol, secoisolaricresinol, lariciresinol, pinoresinol, syringaresinol, medioresinol	Time-resolved fluoroimmunoassay in Finland	Pearson's correlation	0.22	0.01
Hedelin <i>et al.</i> ⁽¹¹⁾	Swedish men (n 177)	FFQ (261 food items)	Matairesinol, secoisolaricresinol, lariciresinol, pinoresinol, syringaresinol, medioresinol	Time-resolved fluoroimmunoassay in Finland	Pearson's correlation	0.19	0.09
Bhakta <i>et al.</i> ⁽²⁸⁾	South Asian women in UK (n 58)	FFQ (277 food items) and 24 h recall	Matairesinol, secoisolaricresinol	Time-resolved fluoroimmunoassay in Finland	Spearman's correlation	0.1 for FFQ; 0.08 for 24 h recall	0.43 for FFQ; 0.5 for 24 h recall
Bhakta <i>et al.</i> ⁽²⁷⁾	South Asian women in UK (n 100); native British women (n 40)	24 h recall	Matairesinol, secoisolaricresinol	Time-resolved fluoroimmunoassay in Finland	Spearman's correlation	r 0.1 for South Asian women; 0.08 for native British	0.2 for South Asian women; 0.6 for native British
Kilkinen <i>et al.</i> ⁽²⁶⁾	Finnish men and women (n 1784)	24 h recall	Matairesinol, secoisolaricresinol	Time-resolved fluoroimmunoassay in Finland	Spearman's correlation	0.19	< 0.0001

the FFQ were not designed specifically for lignan intake; thus, it was difficult to obtain full coverage of related food sources. On the other hand, concentration of enterolactone might vary substantially over the course of a single day, or different seasons. Furthermore, potentially large inter-individual variation in the metabolism of plant lignans into mammalian enterolactone cannot be ignored⁽⁵⁴⁾. Upon ingestion, plant lignans are transformed into their immediate precursor enterodiols and then to enterolactone by certain gut microflora^(4,5,39). Therefore, lack of certain gut microflora can result in different enterodiols:enterolactone ratios. Moreover, persons regularly consuming certain lignan-rich foods, such as fruits, vegetables and fibre-rich foods have more efficient transformation of plant lignans into mammalian enterolactone^(55,56). Smoking and high BMI might decrease the serum concentration of enterolactone, while constipation might enhance the production of enterolactone due to the decrease of intestinal motility^(44,57).

To improve the assessment of phyto-oestrogens using FFQ, some modifications might be warranted. For example, some improvements might be needed to develop a lignan-specific questionnaire, in which important exposures such as age, education, height and weight (to calculate BMI) will also be included. Furthermore, several criteria should be considered when adopting a biomarker to calibrate dietary assessment, e.g. by obtaining serum samples at several time points. Although hampered by low statistical power, it was interesting that obese participants seemed to show a higher correlation between estimate of plant lignan intake and serum concentration of enterolactone. Speculatively, obese people might be more prone to recall healthy dietary habits, such as intake of fruits and vegetables, compared to non-obese people. The difference in assessment of dietary lignan intake using the FFQ-97 and the FFQ-87 might be due to the fact that the FFQ-97 (sixty-five food items containing lignans) evaluated more lignan-containing food items than the FFQ-87 (forty-five food items containing lignans), and the fact that some questions in the FFQ-97 were more precise than those in the FFQ-87. Participants were asked to report food intake frequency within eight categories ranging from 'never/seldom' to 'four times/d' in the FFQ-87, whereas they were asked about the precise frequency (open answers, not pre-specified categories) for commonly consumed food items, e.g. tea, in the FFQ-97. In fact, there were differences between the FFQ-87 and the FFQ-97 in terms of assessment of tea.

In summary, the present study indicates that the correlation between plant lignan intake based on assessment with the FFQ and serum concentration of enterolactone is limited, which can partly depend on varying metabolism of lignans. Therefore, interpretation of FFQ-based results regarding lignan exposure should be very cautious, and direct measurement of serum enterolactone should be recommended.

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References

- Adlercreutz H, Bannwart C, Wahala K, *et al.* (1993) Inhibition of human aromatase by mammalian lignans and isoflavonoid phytoestrogens. *J Steroid Biochem Mol Biol* **44**, 147–153.
- Adlercreutz H & Mazur W (1997) Phyto-oestrogens and Western diseases. *Ann Med* **29**, 95–120.
- Adlercreutz H (2007) Lignans and human health. *Crit Rev Clin Lab Sci* **44**, 483–525.
- Borriello SP, Setchell KD, Axelson M, *et al.* (1985) Production and metabolism of lignans by the human faecal flora. *J Appl Bacteriol* **58**, 37–43.
- Setchell KD, Lawson AM, Borriello SP, *et al.* (1981) Lignan formation in man—microbial involvement and possible roles in relation to cancer. *Lancet* **ii**, 4–7.
- Vanharanta M, Voutilainen S, Lakka TA, *et al.* (1999) Risk of acute coronary events according to serum concentrations of enterolactone: a prospective population-based case–control study. *Lancet* **354**, 2112–2115.
- Reynolds K, Chin A, Lees KA, *et al.* (2006) A meta-analysis of the effect of soy protein supplementation on serum lipids. *Am J Cardiol* **98**, 633–640.
- Yamamoto S, Sobue T, Kobayashi M, *et al.* (2003) Soy, isoflavones, and breast cancer risk in Japan. *J Natl Cancer Inst* **95**, 906–913.
- Shu XO, Jin F, Dai Q, *et al.* (2001) Soyfood intake during adolescence and subsequent risk of breast cancer among Chinese women. *Cancer Epidemiol Biomarkers Prev* **10**, 483–488.
- Pietinen P, Stumpf K, Mannisto S, *et al.* (2001) Serum enterolactone and risk of breast cancer: a case–control study in eastern Finland. *Cancer Epidemiol Biomarkers Prev* **10**, 339–344.
- Hedelin M, Klint A, Chang ET, *et al.* (2006) Dietary phytoestrogen, serum enterolactone and risk of prostate cancer: the Cancer Prostate Sweden Study (Sweden). *Cancer Cause Control* **17**, 169–180.
- Strom SS, Yamamura Y, Duphorne CM, *et al.* (1999) Phytoestrogen intake and prostate cancer: a case–control study using a new database. *Nutr Cancer J* **33**, 20–25.
- Severson RK, Nomura AM, Grove JS, *et al.* (1989) A prospective study of demographics, diet, and prostate cancer among men of Japanese ancestry in Hawaii. *Cancer Res* **49**, 1857–1860.
- Nagata C, Takatsuka N, Kawakami N, *et al.* (2001) Soy product intake and hot flashes in Japanese women: results from a community-based prospective study. *Am J Epidemiol* **153**, 790–793.
- Albertazzi P, Pansini F, Bonaccorsi G, *et al.* (1998) The effect of dietary soy supplementation on hot flashes. *Obstet Gynecol* **91**, 6–11.
- Murkies AL, Lombard C, Strauss BJ, *et al.* (1995) Dietary flour supplementation decreases post-menopausal hot flashes: effect of soy and wheat. *Maturitas* **21**, 189–195.
- Hedelin M, Lof M, Olsson M, *et al.* (2008) Dietary phytoestrogens are not associated with risk of overall breast cancer but diets rich in coumestrol are inversely associated with risk of estrogen receptor and progesterone receptor negative breast tumors in Swedish women. *J Nutr* **138**, 938–945.
- Stattin P, Adlercreutz H, Tenkanen L, *et al.* (2002) Circulating enterolactone and prostate cancer risk: a Nordic nested case-control study. *Int J Cancer* **99**, 124–129.
- Keinan-Boker L, van Der Schouw YT, Grobbee DE, *et al.* (2004) Dietary phytoestrogens and breast cancer risk. *Am J Clin Nutr* **79**, 282–288.
- Horn-Ross PL, Hoggatt KJ, West DW, *et al.* (2002) Recent diet and breast cancer risk: the California Teachers Study (USA). *Cancer Causes Control* **13**, 407–415.
- Key TJ, Sharp GB, Appleby PN, *et al.* (1999) Soya foods and breast cancer risk: a prospective study in Hiroshima and Nagasaki, Japan. *Br J Cancer* **81**, 1248–1256.
- Lee HP, Gourley L, Duffy SW, *et al.* (1991) Dietary effects on breast-cancer risk in Singapore. *Lancet* **337**, 1197–1200.
- den Tonkelaar I, Keinan-Boker L, Veer PV, *et al.* (2001) Urinary phytoestrogens and postmenopausal breast cancer risk. *Cancer Epidemiol Biomarkers Prev* **10**, 223–228.
- dos Santos Silva I, Mangtani P, McCormack V, *et al.* (2004) Phyto-oestrogen intake and breast cancer risk in South Asian women in England: findings from a population-based case–control study. *Cancer Causes Control* **15**, 805–818.
- Willett WC (1998) Correction for the effects of measurement error. In *Nutritional Epidemiology*, chapter 2, pp. 302–320 [WC Willett, editor]. Oxford: University Press.
- Kilkinen A, Valsta LM, Virtamo J, *et al.* (2003) Intake of lignans is associated with serum enterolactone concentration in Finnish men and women. *J Nutr* **133**, 1830–1833.
- Bhakta D, Higgins CD, Sevak L, *et al.* (2006) Phyto-oestrogen intake and plasma concentrations in South Asian and native British women resident in England. *Br J Nutr* **95**, 1150–1158.
- Bhakta D, dos Santos Silva I, Higgins C, *et al.* (2005) A semi-quantitative food frequency questionnaire is a valid indicator of the usual intake of phytoestrogens by south Asian women in the UK relative to multiple 24-h dietary recalls and multiple plasma samples. *J Nutr* **135**, 116–123.
- Adlercreutz H, Fotsis T, Bannwart C, *et al.* (1986) Determination of urinary lignans and phytoestrogen metabolites, potential antiestrogens and anticarcinogens, in urine of women on various habitual diets. *J Steroid Biochem* **25**, 791–797.
- Bergström L, Kylberg E, Hagman U, *et al.* (1991) The food composition database KOST: the National Food Administration's information system for nutritive values of food. *Vår Föda* **43**, 439–447.
- Milder IEJ, Arts ICW, van de Putte B, *et al.* (2005) Lignan contents of Dutch plant foods: a database including lariciresinol, pinoresinol, secoisolariciresinol and matairesinol. *Br J Nutr* **93**, 393–402.
- Valsta LM, Kilkinen A, Mazur W, *et al.* (2003) Phyto-oestrogen database of foods and average intake in Finland. *Br J Nutr* **89**, S31–S38.
- Mazur W & Adlercreutz H (1998) Natural and anthropogenic environmental oestrogens: the scientific basis for risk assessment. Naturally occurring oestrogens in food. *Pure Appl Chem* **70**, 1759–1776.
- Mazur WM, Uehara M, Wahala K, *et al.* (2000) Phyto-oestrogen content of berries, and plasma concentrations and urinary excretion of enterolactone after a single strawberry-meal in human subjects. *Br J Nutr* **83**, 381–387.

35. Mazur WM, Duke JA, Wahala K, *et al.* (1998) Isoflavonoids and lignans in legumes: nutritional and health aspects in humans. *J Nutr Biochem* **9**, 193–200.
36. Mazur WM, Wahala K, Rasku S, *et al.* (1998) Lignan and isoflavonoid concentrations in tea and coffee. *Br J Nutr* **79**, 37–45.
37. Mazur W, Fotsis T, Wahala K, *et al.* (1996) Isotope dilution gas chromatographic–mass spectrometric method for the determination of isoflavonoids, coumestrol, and lignans in food samples. *Anal Biochem* **233**, 169–180.
38. Willett W & Stampfer MJ (1986) Total energy intake: implications for epidemiologic analyses. *Am J Epidemiol* **124**, 17–27.
39. Setchell K & Adlercreutz H (1998) Mammalian lignans and phytoestrogens: recent studies on their formation, metabolism and biological roles in health and disease. In *The Role of Gut Microflora in Toxicity and Cancer*, pp. 315–345 [IR Rowland, editor]. London: Academic Press.
40. Heinonen S, Nurmi T, Liukkonen K, *et al.* (2001) *In vitro* metabolism of plant lignans: new precursors of mammalian lignans enterolactone and enterodiol. *J Agric Food Chem* **49**, 3178–3186.
41. Wang GJ, Lapcik O, Hampl R, *et al.* (2000) Time-resolved fluoroimmunoassay of plasma daidzein and genistein. *Steroids* **65**, 339–348.
42. Adlercreutz H, Wang GJ, Lapcik O, *et al.* (1998) Time-resolved fluoroimmunoassay for plasma enterolactone. *Anal Biochem* **265**, 208–215.
43. Stumpf K, Uehara M, Nurmi T, *et al.* (2000) Changes in the time-resolved fluoroimmunoassay of plasma enterolactone. *Anal Biochem* **284**, 153–157.
44. Kilkkinen A, Stumpf K, Pietinen P, *et al.* (2001) Determinants of serum enterolactone concentration. *Am J Clin Nutr* **73**, 1094–1100.
45. Heald CL, Bolton-Smith C, Ritchie MR, *et al.* (2006) Phytoestrogen intake in Scottish men: use of serum to validate a self-administered food-frequency questionnaire in older men. *Eur J Clin Nutr* **60**, 129–135.
46. Sonestedt E & Wirfalt E (2010) Enterolactone and breast cancer: methodological issues may contribute to conflicting results in observational studies. *Nutr Res* **30**, 667–677.
47. Sonestedt E, Ericson U, Gullberg B, *et al.* (2008) Variation in fasting and non-fasting serum enterolactone concentrations in women of the Malmo Diet and Cancer cohort. *Eur J Clin Nutr* **62**, 1005–1009.
48. Hulten K, Winkvist A, Lenner P, *et al.* (2002) An incident case-referent study on plasma enterolactone and breast cancer risk. *Eur J Nutr* **41**, 168–176.
49. Barnes S, Sfakianos J, Coward L, *et al.* (1996) Soy isoflavonoids and cancer prevention. Underlying biochemical and pharmacological issues. *Adv Exp Med Biol* **401**, 87–100.
50. Leiter EH, Chapman HD & Falany CN (1991) Synergism of obesity genes with hepatic steroid sulfotransferases to mediate diabetes in mice. *Diabetes* **40**, 1360–1363.
51. Takaishi H, Matsuki T, Nakazawa A, *et al.* (2008) Imbalance in intestinal microflora constitution could be involved in the pathogenesis of inflammatory bowel disease. *Int J Med Microbiol* **298**, 463–472.
52. Kilkkinen A, Pietinen P, Klaukka T, *et al.* (2002) Use of oral antimicrobials decreases serum enterolactone concentration. *Am J Epidemiol* **155**, 472–477.
53. Hausner H, Johnsen NF, Hallund J, *et al.* (2004) A single measurement is inadequate to estimate enterolactone levels in Danish postmenopausal women due to large intraindividual variation. *J Nutr* **134**, 1197–1200.
54. Rowland IR, Wiseman H, Sanders TA, *et al.* (2000) Interindividual variation in metabolism of soy isoflavones and lignans: influence of habitual diet on equol production by the gut microflora. *Nutr Cancer* **36**, 27–32.
55. Nurmi T, Mursu J, Penälvo JL, *et al.* (2010) Dietary intake and urinary excretion of lignans in Finnish men. *Br J Nutr* **103**, 677–685.
56. Hutchins AM, Lampe JW, Martini MC, *et al.* (1995) Vegetables, fruits, and legumes: effect on urinary isoflavonoid phytoestrogen and lignan excretion. *J Am Diet Assoc* **95**, 769–774.
57. Johnsen NF, Hausner H, Olsen A, *et al.* (2004) Intake of whole grains and vegetables determines the plasma enterolactone concentration of Danish women. *J Nutr* **134**, 2691–2697.