# VIM- and IMP-Type Metallo-β-lactamase Producing *Pseudomonas* spp. and *Acinetobacter* spp. in Korean Hospitals

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We determined the occurrence of acquired metallo- $\beta$ -lactamase (MBL)–producing bacteria in Korean hospitals. Among the isolates nonsusceptible to imipenem that were collected from 28 hospitals from 2000 to 2001, 44 (11.4%) of 387 *Pseudomonas* spp. and 38 (14.2%) of 267 *Acinetobacter* spp. produced MBL and had alleles of *bla*<sub>VIM-2</sub> or *bla*<sub>IMP-1</sub>. MBL-producing isolates were detected in 60.7% of the hospitals.

Carbapenems are often used as a last resort for treating serious infections attributable to multidrug-resistant gram-negative bacilli because these drugs are stable even to extended-spectrum and AmpC  $\beta$ -lactamases. However, gram-negative bacilli with acquired metallo- $\beta$ -lactamase (MBL), IMP-1, emerged and spread during the early 1990s in Japan (1). IMP-1 and its variants were then detected in other countries (2).

Another type of acquired MBL, VIM-1, was first reported in *Pseudomonas aeruginosa* in Italy (3), followed by reports of VIM-2 in France and Greece. VIM-2 was detected in *P. aeruginosa* in a Korean hospital isolated as early as 1995 (4). The occurrence of the VIM enzyme has continued to evolve: VIM-3 was reported in Taiwan (5), and VIM-4 in the United States (6).

The  $bla_{\rm IMP}$  and  $bla_{\rm VIM}$  genes are horizontally transferable because they are inserted in integrons, and some of these integrons are located on conjugative plasmids (7). Because of its ability to spread, carbapenem resistance related to IMP and VIM  $\beta$ -lactamase production has become a serious concern (8). Laboratory personnel and physicians must consider the therapeutic and infectioncontrol implications of not detecting carbapenemase-producing bacteria (9). A large number of VIM-2-producing Pseudomonas spp. have been detected in a Korean hospital since 1995 (4), but the occurrence of MBL-producing isolates has not been studied at other Korean hospitals, despite the high prevalence of carbapenem-resistant P. aeruginosa and Acinetobacter spp. (10). The aim of our study was to determine the occurrence of acquired MBLproducing P. aeruginosa and Acinetobacter spp. among isolates collected by Korean Nationwide Surveillance of Antimicrobial Resistance Group hospitals. The MBL types produced and the sources of the MBL-positive isolates were also investigated. In addition, pulsed-field gel electrophoresis (PFGE) patterns were compared to determine intra- and inter-hospital spread of resistant strains.

### The Study

Nonduplicate, imipenem-resistant isolates of 387 *Pseudomonas* spp. and 267 *Acinetobacter* spp. were collected from 2000 to 2001 from 28 hospitals in the Korean Nationwide Surveillance of Antimicrobial Resistance Group hospitals located in six cities or provinces. The identification of the species and the imipenem susceptibility were confirmed at the coordinating laboratory by using conventional tests (11) or ATB 32 GN system (bioMerieux, Marcy-l'Etoile, France) and by using the disk diffusion test (12), respectively.

MBL production was screened by using the Hodge test and the imipenem-EDTA double disk synergy test (13). The  $bla_{IMP-1}$  and  $bla_{VIM-2}$  alleles were detected by polymerase chain reaction (PCR), and three of the positive isolates were confirmed by sequencing, as described previously (4). *Xba*I-digested genomic DNA of *P. aeruginosa* isolates was separated by PFGE using the CHEF-DR-II system (Bio-Rad Laboratories, Hercules, CA) (4). The pattern was analyzed visually and by using UVIBand and Map software (UVItec Ltd., Cambridge, UK).

Some of the *Pseudomonas* and *Acinetobacter* isolates collected were not fully resistant to imipenem but showed intermediate resistance when retested. Among the isolates not susceptible to imipenem, 44 (11.4%) of 387 *Pseudomonas* spp. (42 *P. aeruginosa* and 2 *P. putida*) and 38 (14.2%) of 267 *Acinetobacter* spp. were considered MBL producers on the basis of positive results by the

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		No. hos	spitals (%)	No. isolates (%)	
Organism	City/province	Tested	Positive	Tested	Positive
Pseudomonas spp.	Seoul	11 <sup>a</sup>	4 (36.4)	144	12 (8.3)
	Kyungki	2	2 (100)	40	6 (15.0)
	Kangwon	2	1 (50.0)	57	2 (3.5)
	Chulla	4	4 (100)	108	24 (22.2)
	Kyungsang	2	0 (0)	38	0 (0)
	Total	21	11 (52.4)	387	44 (11.4)
Acinetobacter spp.	Seoul	11 <sup>a</sup>	4 (36.4)	107	12 (11.2)
	Kyungki	3	0 (0)	29	0 (0)
	Kangwon	3	2 (25.0)	41	8 (19.5)
	Chulla	3	1 (12.5)	25	13 (52.0)
	Kyungsang	2	1 (50.0)	53	1 (1.9)
	Chungchung	2	2 (100)	12	4 (33.3)
	Total	24	10 (41.7)	267	38 (14.2)
<sup>a</sup> Four were tertiary-care hospitals.					

Table 1. Detection of metallo- $\beta$ -lactamase-producing isolates among imipenem-nonsusceptible isolates of *Pseudomonas* spp. and *Acinetobacter* spp.

Hodge test and imipenem-EDTA double disk synergy test (Table 1). MBL-producing *Pseudomonas* spp. and *Acinetobacter* spp. were detected in 11 (52.4%) of 21 and 10 (41.7%) of 24 hospitals that were located in four of five and five of six cities or provinces, respectively. We detected the *bla*<sub>VIM</sub> allele by PCR from all 42 isolates of MBL-producing *P. aeruginosa* and 2 isolates of *P. putida*. The *bla*<sub>VIM-2</sub> and *bla*<sub>IMP-1</sub> alleles were detected in 27 (71.1%) and 11 (28.9%) of 38 *Acinetobacter* isolates, respectively (Table 2). Nucleotide sequencing for three representative PCR-positive isolates confirmed the presence of the *bla*<sub>VIM-2</sub> gene in one isolate each of *P. aeruginosa* and *Acinetobacter* spp., and the *bla*<sub>IMP-1</sub> gene in one isolate of *Acinetobacter* spp.

The MBL-producing strains were isolated mainly from intensive-care unit patients (31.7%) and other inpatients (50.0%); five (6.1%) were from emergency service and other outpatients (Table 3). Overall, MBL-producing isolates were mainly obtained from specimens of sputum (50.0%) and urine (29.3%). However, the proportion of MBL-producing isolates was relatively higher among urine isolates: 17.3% for *Pseudomonas* spp. and 29.2% for *Acinetobacter* spp. We obtained one MBL-producing *Acinetobacter* isolate from each of the following specimen types: blood, spinal fluid, pleural fluid, and venous catheter tip (Table 4).

Table 2. Detection of <i>bla</i> <sub>VIM-2</sub> and <i>bla</i> <sub>IMP-1</sub> allele from metallo-β-
lactamase-producing Pseudomonas spp. and Acinetobacter
spp. by polymerase chain reaction

	No. isolates (%)			
Organism	Tested	<i>bla</i> <sub>VIM-2</sub> positive	<i>bla</i> <sub>IMP-1</sub> positive	
Pseudomonas aeruginosa	42	42 (100)	0 (0)	
P. putida	2	2 (100)	0 (0)	
Acinetobacter spp.	38	27 (71.1)	11 (28.9)	
Total	82	71 (86.6)	11 (13.4)	

The PFGE of the *Xba*I-digested genomic DNA of 39 isolates of *P. aeruginosa* showed 22 patterns (data not shown). Six isolates from one hospital had an identical pattern. Thirteen isolates (33.3%) belonged to another identical pattern—six from one hospital, two from each of two hospitals, and one from each of three hospitals, which were located in a city and two provinces.

#### Conclusions

In this study, >10% of all imipenem-nonsusceptible isolates of *Pseudomonas* spp. and *Acinetobacter* spp. were attributable to MBL production (Table 1), and these MBLproducing isolates were detected in 62.5% of the participating hospitals. Our finding indicates that MBL-producing *P. aeruginosa* is more prevalent in Korea than in other countries (2) and that MBL-producing *Acinetobacter* spp. is increasing. The percentage of hospitals with MBL-producing isolates might have been higher if a larger number of imipenem-nonsusceptible isolates had been collected for this study.

VIM-2 was the only type of acquired MBL identified initially in Korea. VIM-2–producing *P. aeruginosa* was isolated at almost the same time in Europe (7) and Korea (4). However, IMP-1–producing isolates were rare until 2000 in Korea. Only one and three IMP-1-positive *P. aeruginosa* and *Acinetobacter* spp., respectively, have been isolated at the coordinating laboratory (4, unpub. data). In our study, 11 (28.9%) of 38 MBL-positive isolates of *Acinetobacter* spp. were IMP producers (Table 2). This increase suggests the possible introduction of IMP-producing strains of *Acinetobacter* spp. from Japan, where 28 isolates of *bla*<sub>IMP-1</sub>-positive *Acinetobacter baumannii* were reported in a hospital as early as 1994 to 1996 (14).

Rasmussen and Bush (15) predicted that an increase of MBL-producing organisms was inevitable, given the more frequent use of carbapenems. Imipenem has been used for

#### DISPATCHES

	No. isolates (%)					
Organism	Outpatient	Inpatient	Intensive-care unit	Others	Total	
Pseudomonas spp.	$3(6.8)^{a}$	26 (59.1)	11 (25.0)	4 (9.1)	44 (100)	
Acinetobacter spp.	2 (5.2) <sup>b</sup>	15 (39.5)	15 (39.5)	6 (15.8)	38 (100)	
Total	5 (6.1)	41 (50.0)	26 (31.7)	10 (12.2)	82 (100)	
<sup>a</sup> Two were emergency service patients, and one was a urology patient.						

Table 3. blavIM-2 and blaIMP-1 allele-positive Pseudomonas spp. and Acinetobacter spp. isolated by service

<sup>b</sup>One was an emergency service patient, and one was a pediatric patient

only 9 years in Korea, but the imipenem-resistance rate of P. aeruginosa has rapidly risen from 6% in 1996 to 19% in 2001. A study by the Korean Nationwide Surveillance of Antimicrobial Resistance Group showed that the mean imipenem-resistance rates of P. aeruginosa in 1997 did not differ substantially depending on hospital size, (i.e., 17% in medium hospitals [<1,000 beds] and 18% in large hospitals [≥1,000 beds]). The mean resistance rates to imipenem were not lower than those to ceftazidime in 2000, i.e., 21% versus 18% in large hospitals and 20% versus 19% in medium hospitals (data not shown).

Acinetobacter spp. are also common nosocomial pathogens with multidrug resistance. The imipenem resistance rate of this organism isolated in Korea was found to be much lower than that of P. aeruginosa, but its resistance rate rose from 4% in the first quarter to 20% in the third quarter of 2002 at the coordinating laboratory (data not shown).

In our study, MBL-producing Pseudomonas spp. and Acinetobacter spp. were isolated mainly from sputum and urine specimens, and most (81.7%) isolates were from inpatients and intensive-care unit patients. Therefore, proper treatment of respiratory secretions and urine from intensive-care unit patients is considered an important aspect of preventing the spread of MBL-producing organisms. The presence of P. aeruginosa isolates with identical PFGE patterns among those collected not only from certain hospitals but also from different hospitals suggests that clonal spread is at least a part of the cause of intra- and inter-hospital dissemination of MBL-producing isolates. The presence of VIM-2-producing Serratia marcescens, Enterobacter cloacae, and Achromobacter xylosoxidans subsp. denitrificans (unpub. data) in other hospitals also suggests horizontal transfer of the resistance determinants.

Cornaglia et al. reported that five of seven patients infected with MBL-producing P. aeruginosa died, although the cause of death was difficult to establish with certainty (16). Clinical studies on the infection are rare because isolation of MBL-producing gram-negative bacilli increased only recently. We anticipate difficulties in treating patients infected with MBL-producing gram-negative bacilli, which can hydrolyze, in vitro, all available  $\beta$ lactams, except aztreonam for which clinical efficacy is unknown.

Our study indicates the urgent need for action to prevent further spread of MBL-producing organisms. Previous experiences with penicillin-nonsusceptible pneumococci, methicillin-resistant Staphylococcus aureus, vancomycin-resistant Enterococcus faecium, and extendedspectrum B-lactamase-producing Klebsiella pneumoniae indicate that once resistant bacteria can become widespread they cannot be controlled (10). Our first task is to detect MBL producers among clinical isolates (9). Although the National Committee for Clinical Laboratory Standards document (12) does not contain procedures for detection, simple procedures are available (13).

The prevalence of *bla*<sub>VIM-2</sub> allele-positive *P. aeruginosa* and *bla*<sub>IMP-1</sub> allele-positive *Acinetobacter* spp. is increasing possibly because of clonal and horizontal spread of the resistance determinant in Korean hospitals. Sputum and urine from inpatients and intensive-care unit patients were found to be the main sources of MBL-producing isolates. Laboratories not only in Korea but also in other countries with carbapenem-resistant organisms must be prepared to screen MBL-producing isolates to determine the clinical impact and prevent further spread of MBL-producing organisms.

Table 4. blavim-2 and blaimP-1 allele-positive Pseudomonas spp. and Acinetobacter spp. isolated by source							
	No. (%) of isolates with metallo-â-lactamase						
	Pseudon	<i>ionas</i> spp.	Acinetobacter spp.		Total		% positive by
Source	Tested	Positive	Tested	Positive	Tested	Positive	source
Sputum	200	22 (11.0)	143	19 (13.3)	343	41 (12.0)	50.0
Urine	98	17 (17.3)	24	7 (29.2)	122	24 (19.7)	29.3
Wound	49	2 (4.1)	71	7 (9.9)	120	9 (7.5)	10.9
Other <sup>a</sup>	18	3 (16.7)	29	5 (17.2)	47	8 (17.0)	9.8
Total	387	44 (11.4)	267	38 (14.2)	654	82 (12.5)	100

<sup>a</sup>Others included one Acinetobacter isolate of specimens from blood, spinal fluid, pleural fluid, and a venous catheter tip.

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#### References

- Osano E, Arakawa Y, Wacharotayankun R, Ohta M, Horii T, Ito H, et al. Molecular characterization of an enterobacterial metallo-β-lactamases found in a clinical isolates of *Serratia marcescens* that shows imipenem resistance. Antimicrob Agents Chemother 1994;38:71–8.
- Gibb AP, Tribuddharat C, Moore RA, Louie TJ, Krulicki W, Livermore DM, et al. Nosocomial outbreak of carbapenem-resistant *Pseudomonas aeruginosa* with a new *bla*<sub>IMP</sub> allele, *bla*<sub>IMP-7</sub>. Antimicrob Agents Chemother 2002;46:255–8.
- Lauretti L, Riccio ML, Mazzariol A, Cornaglia G, Amicosante G, Fontana R, et al. Cloning and characterization of *bla*<sub>VIM</sub>, a new integron-borne metallo-β-lactamase gene from a *Pseudomonas aeruginosa* clinical isolate. Antimicrob Agents Chemother 1999; 43:1584–90.
- Lee K, Lim JB, Yum JH, Yong D, Chong Y, Kim JM, et al. *bla*<sub>VIM-2</sub> cassette-containing novel integrons in metallo-β-lactamase–producing *Pseudomonas aeruginosa* and *Pseudomonas putida* isolates disseminated in a Korean hospital. Antimicrob Agents Chemother 2002;46:1053–8.
- Yan J-J, Hsueh P-R, Ko W-C, Luh E-T, Tsai S-H, Wu H-M, et al. Metallo-β-lactamases in clinical *Pseudomonas* isolates in Taiwan and identification of VIM-3, a novel variant of the VIM-2 enzyme. Antimicrob Agents Chemother 2001;45:2224–8.
- Tolman MA, Rolston K, Jones RN, Walsh TR. Molecular characterization of VIM-4, a novel metallo-beta-lactamase isolated from Texas: report from the cancer surveillance program (2001). Proceedings of the 42nd Interscience Conference on Antimicrobial Agents and Chemotherapy. Abstract C1-1851. San Diego, California; 2002.

- Poirel L, Naas T, Nicolas D, Collet L, Bellais S, Cavallo T-D, et al. Characterization of VIM-2, a carbapenem-hydrolyzing metallo-β-lactamase and its plasmid- and integron borne gene from a *Pseudomonas aeruginosa* clinical isolate in France. Antimicrob Agents Chemother 2000;44:891–7.
- Richet HM, Mohammed J, McDonald LC, Jarvis WR, INSPEAR. Building communication networks: International Network for the Study and Prevention of Emerging Antimicrobial Resistance. Emerg Infect Dis 2001;7:319–22.
- Molland ES, Black JA, Ourada J, Reisbid MD, Hanson ND, Thomson KS. Occurrence of newer β-lactamases in *Klebsiella pneumoniae* isolates from 24 U.S. hospitals. Antimicrob Agents Chemother 2002;46:3837–42.
- 10. Lee K, Lee HS, Jang SJ, Park AJ, Lee MH, Song WK, et al. Antimicrobial resistance surveillance of bacteria in 1999 in Korea with a special reference to resistance of enterococci to vancomycin and gram-negative bacilli to third generation cephalosporin, imipenem, and fluoroquinolone. J Korean Med Sci 2001;16:262–70.
- Kiska DL, Gilligan PH. *Pseudomonas*. In: Murray PR, Baron EJ, Pfaller MA, Tenover FC, Yolken RH, editors. Manual of clinical microbiology, 7th ed. Washington: American Society for Microbiology; 1999. p. 517–25.
- National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial susceptibility testing. Eleventh informational supplement. NCCLS document M100-S11. Wayne (PA): The Committee, 2001.
- Lee K, Chong Y, Shin HB, Kim YA, Yong D, Yum JH. Modified Hodge test and EDTA-disk synergy tests to screen metallo-β-lactamase-producing strains of *Pseudomonas* and *Acinetobacter* species. Clin Microbiol Infect 2001;7:88–91.
- Takahashi A, Yomoda S, Kobayashi I, Kubo T, Tsunoda M, Iyobe S. Detection of carbapenemase-producing *Acinetobacter baumannii* in a hospital. J Clin Microbiol 2000;38:526–9.
- Rasmussen BA, Bush K. Carbapenem-hydrolyzing β-lactamases. Antimicrob Agents Chemother 1997;41:223–32.
- Cornaglia G, Mazzariol A, Lauretti L, Rossolini GM, Fontana R. Hospital outbreak of carbapenem-resistant *Pseudomonas aeruginosa* producing VIM-1, a novel transferable metallo-β-lactamase. Clin Infect Dis 2000;31:1119–25.

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