

## 70th Anniversary Conference on ‘From plough through practice to policy’

### Symposium 3: Obesity-related cancers Visceral obesity, metabolic syndrome, insulin resistance and cancer

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This paper presents emerging evidence linking visceral adiposity and the metabolic syndrome (MetSyn) with carcinogenesis. The link between obesity and cancer has been clearly identified in a multitude of robust epidemiological studies. Research is now focusing on the role of visceral adipose tissue in carcinogenesis; as it is recognised as an important metabolic tissue that secretes factors that systemically alter the immunological, metabolic and endocrine milieu. Excess visceral adipose tissue gives rise to a state of chronic systemic inflammation with associated insulin resistance and dysmetabolism, collectively known as the MetSyn. Prospective cohort studies have shown associations between visceral adiposity, the MetSyn and increased risk of breast cancer, colorectal cancer and oesophageal adenocarcinoma. Furthermore, visceral adiposity and the MetSyn have been associated with increased tumour progression and reduced survival. The mechanisms by which visceral adiposity and the MetSyn are thought to promote tumorigenesis are manifold. These include alterations in adipokine secretion and cell signalling pathways. In addition, hyperinsulinaemia, subsequent insulin resistance and stimulation of the insulin-like growth factor-1 axis have all been linked with visceral adiposity and promote tumour progression. Furthermore, the abundance of inflammatory cells in visceral adipose tissue, including macrophages and T-cells, create systemic inflammation and a pro-tumorigenic environment. It is clear from current research that excess visceral adiposity and associated dysmetabolism play a central role in the pathogenesis of certain cancer types. Further research is required to elucidate the exact mechanisms at play and identify potential targets for intervention.

#### Visceral adiposity: Metabolic syndrome: Insulin resistance: Inflammation: Tumorigenesis

The prevalence of obesity has increased rapidly in recent years<sup>(1)</sup>. Being overweight or obese is now the most prevalent body composition in some countries, accounting for 71% of males and 62% of females in the United States<sup>(2)</sup>. Similar trends exist in Western Europe with greater than 66% of men and 49–57% of women being overweight or obese in the UK and Ireland<sup>(3,4)</sup>. This pattern shows no signs of abating as obesity rates are increasing among children<sup>(5)</sup> and overweight children tend to become overweight adults<sup>(6)</sup>. In countries where sedentary lifestyles and high-energy foods are abundant it is easy to see how energy intake exceeds that expended. It has been estimated that ingestion of 5% more energy than expended may

result in an accumulation of 5 kg adipose tissue in a single year<sup>(7)</sup>.

Epidemiological studies have demonstrated a robust link between obesity and cancer development at numerous sites, in particular the oesophagus, pancreas, colorectum, breast (postmenopausal), endometrium and kidney<sup>(8,9)</sup>. This association carries relative risk (RR) estimates of 1.1–1.6 per 5 kg/m<sup>2</sup> incremental increase in BMI<sup>(8)</sup>. Obesity also increases cancer-related mortality with studies reporting that obesity could account for 14% of all deaths from cancer in men and 20% in women<sup>(10)</sup>. The causative link between obesity and cancer is further strengthened by research in animal models which demonstrates that energy

**Abbreviations:** IGF, insulin-like growth factor; MetSyn, metabolic syndrome; PI3K, phosphoinositide 3-kinase; RR, relative risk.

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restriction decreases spontaneous tumour occurrence<sup>(11,12)</sup>. Furthermore, emerging clinical research suggests weight loss following bariatric surgery leads to a reduction in cancer incidence<sup>(13)</sup>. Thus, the potential mechanisms by which obesity increases both the incidence of certain malignancies, and cancer deaths, have become the focus of considerable research interest in recent years.

### Adipose tissue and the metabolic syndrome

WHO defines obesity as an abnormal or excessive fat accumulation in adipose tissue, to the extent that health is impaired<sup>(1)</sup>. Fat is principally deposited in two compartments; subcutaneously and viscerally. Attention was first drawn to the different patterns of fat distribution over 60 years ago by Jean Vague. He described two distinct types of fat deposition; upper-body, male-type fat (visceral) and lower-body, female-type fat (subcutaneous), and the association of visceral fat with type 2 diabetes, atherosclerosis and gout<sup>(14,15)</sup>. Visceral adipose tissue largely comprises omental adipose tissue and also includes other intra-abdominal fat sources such as mesenteric fat and is more metabolically active than subcutaneous adipose tissue<sup>(16)</sup>. Visceral adipose tissue has multiple endocrine, metabolic and immunological functions and has been shown to be central to the pathogenesis of the metabolic syndrome (MetSyn), a pro-inflammatory, pro-coagulant state associated with insulin resistance<sup>(17)</sup>. The multiple risk factors that commonly appear together as the MetSyn include abdominal obesity, atherogenic dyslipidaemia (raised TAG and reduced HDL cholesterol), elevated fasting plasma glucose and hypertension<sup>(18)</sup>. The importance of adipose tissue location in terms of dysmetabolism risk is evident as an increased ratio of visceral fat area to subcutaneous fat area has been shown to be strongly related to disorders of glucose and lipid metabolism in obese subjects<sup>(19)</sup>. Furthermore, visceral obesity is more strongly associated with increased risk of insulin resistance, the MetSyn and CVD than BMI alone<sup>(20)</sup>.

Visceral fat has been identified as an independent risk factor for breast cancer<sup>(21)</sup>, oesophageal adenocarcinoma<sup>(22)</sup>, colorectal adenocarcinoma<sup>(23)</sup> and colorectal adenomas<sup>(24,25)</sup>. The other elements of the MetSyn, i.e. dyslipidemia, hypertension and insulin resistance have also been independently linked with increased cancer risk<sup>(26–29)</sup>. The synergistic impact of these factors on cancer risk is the focus of the Me-Can Study, a prospective international population-based study of 580 000 people<sup>(30)</sup>. Initial findings suggest that a combination of components of the MetSyn is associated with increased RR of colorectal cancer development (men: RR 1.25, 95% CI 1.18, 1.32; women: RR 1.14, 95% CI 1.02, 1.18)<sup>(31)</sup>, endometrial cancer (RR 1.37, 95% CI 1.28, 1.46)<sup>(32)</sup>, bladder cancer in men (RR: 1.1, 95% CI 1.01, 1.18)<sup>(33)</sup> and pancreatic cancer in women (RR 1.58, 95% CI 1.34, 1.87)<sup>(34)</sup>. Similarly, in studies of pancreatic and colorectal cancer a high prevalence of MetSyn, central obesity and systemic inflammation has been identified<sup>(35–37)</sup> and has been shown to be associated with advanced tumour stage and reduced survival<sup>(38–41)</sup>.

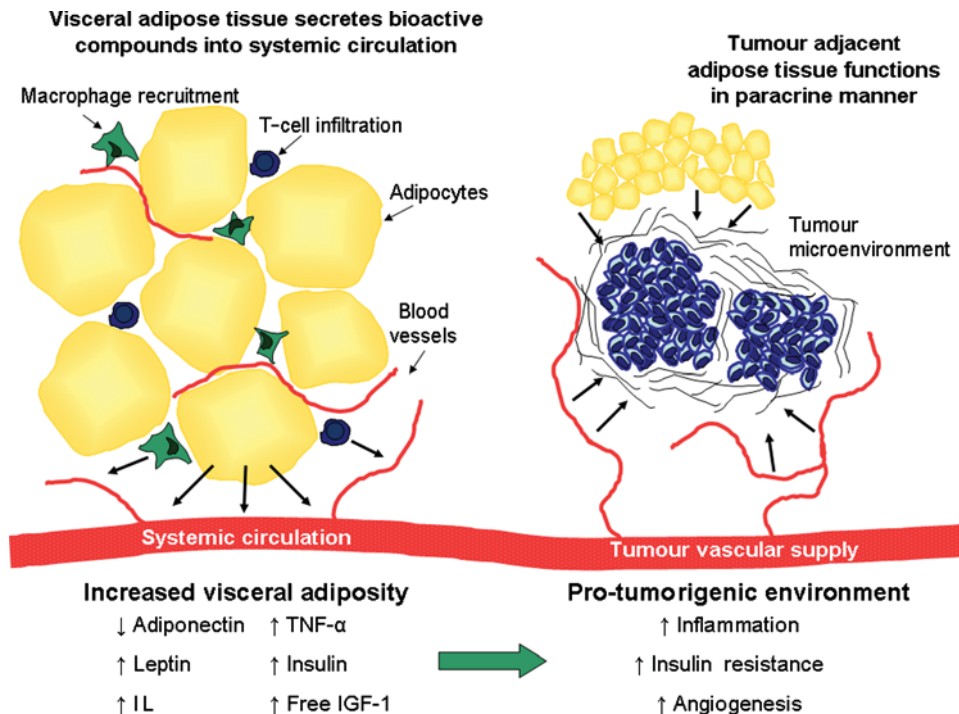
### Oesophageal adenocarcinoma: a model of visceral obesity, metabolic syndrome and cancer

The link between visceral adiposity, the MetSyn and cancer is probably best illustrated in the oesophageal adenocarcinoma model. Oesophageal adenocarcinoma develops along a predefined sequence of events. It begins with the development of Barrett's oesophagus in response to chronic reflux of acid and bile, where the normal squamous lining of the oesophagus is replaced with specialised intestinal metaplasia, which then progresses through to dysplasia and ultimately adenocarcinoma<sup>(42)</sup>. The incidence of oesophageal adenocarcinoma in Ireland has increased by nearly 50% in recent years<sup>(43)</sup>, a trend mirrored across developed countries<sup>(44)</sup> and coincides with the sharp increase in the prevalence of obesity<sup>(4)</sup>. Epidemiological studies highlight a marked association between oesophageal adenocarcinoma and obesity, and a recent meta-analysis reported that a BMI  $\geq 30$  kg/m<sup>2</sup> is associated with a RR of 3.0 for developing oesophageal adenocarcinoma, a higher association than for any other cancer<sup>(45)</sup>.

Visceral obesity itself may promote gastro-oesophageal reflux disease and predispose to Barrett's oesophagus, and this may be one contributory factor to the association with oesophageal adenocarcinoma<sup>(46)</sup>. However, the observation that obesity is a risk factor independent of reflux<sup>(47,48)</sup> suggests other mechanisms are at play. A study at this centre in 2008 reported that the prevalence of the MetSyn in Barrett's oesophagus patients far exceeded population norms with the MetSyn present in 46% of patients and central obesity in 78%. The syndrome was also significantly associated with the length of specialised intestinal metaplasia; 60% of patients with long-segment Barrett's oesophagus had MetSyn and 92% were centrally obese compared with 23.8% and 62% ( $P=0.007$  and  $P=0.005$ ) respectively in short-segment Barrett's oesophagus. Long-segment Barrett's oesophagus was also associated with systemic inflammation, with significantly higher serum levels of IL-6 observed in patients ( $P=0.03$ )<sup>(49)</sup>. The obesity, inflammation, malignancy association is further strengthened by studies which have identified increased visceral adiposity and altered secretion of adipose-derived hormones in oesophageal adenocarcinoma<sup>(22,50)</sup>.

### Mechanisms underlying obesity and tumorigenesis

The mechanisms by which visceral adiposity and the MetSyn are thought to promote tumorigenesis are manifold. Adipose tissue may promote tumour development in a paracrine manner (Fig. 1). This is supported by the observation that epithelial tumour cell growth is enhanced by injection into fat pads rather than subcutaneously<sup>(51)</sup> suggesting that chemokine production within the adipose tissue provides conditions which enhance tumour cell growth. Furthermore, a proteomic study of mammary fat revealed the production of a wide variety of proteins involved in diverse processes such as cell communication, growth, immune response, apoptosis and numerous signalling molecules including hormones, cytokines and growth factors<sup>(52)</sup>. The paracrine influence of adipose



**Fig. 1.** (Colour online) Increased visceral adiposity creates a pro-tumorigenic environment. As visceral adipose tissue expands it becomes infiltrated with macrophages and T-cells. These immune cells and the adipocytes produce adipokines including leptin, IL and TNF- $\alpha$ . In addition, increasing visceral adiposity leads to hyperinsulinaemia and increased levels of free insulin-like growth factor-1 (IGF-1). Consequently, there is a pro-tumorigenic state of inflammation, angiogenesis and insulin resistance. The tumour microenvironment is influenced by adipose-derived factors secreted into systemic circulation and also via paracrine effects of tumour adjacent adipose tissue.

tissue on tumorigenesis is less fully investigated, whereas much research has focused on the systemic effects of visceral adiposity that are putatively involved in cancer biology. These include alterations in adipokine production, insulin and insulin-like growth factor (IGF) pathways, cancer cell signalling and inflammatory pathways (Fig. 1) and are discussed in further detail below.

### Altered adipokine production

The existence of adipose tissue-derived factors that may influence metabolism has been suggested since the 1980s<sup>(53,54)</sup>; however, it was the discovery of the adipokines leptin<sup>(55)</sup> and adiponectin<sup>(56)</sup> which led to the concept of adipose tissue as an endocrine organ. The altered secretion of these and other adipokines in obesity has been implicated in the pathogenesis of obesity-associated pathologies, including cancer<sup>(57)</sup>. Excess adiposity, in particular visceral obesity, results in a state of chronic systemic low-grade inflammation, attributed to production of these inflammatory cytokines by both adipocytes and infiltrating immune cells creating a pro-tumorigenic environment<sup>(58)</sup>.

Adiponectin is the most abundant adipokine and is secreted mainly from adipocytes in visceral fat; however, circulating levels are inversely correlated with obesity<sup>(59)</sup>. It has been shown to be both anti-angiogenic and anti-inflammatory, can inhibit tumour growth in animal models

and circulating levels are lower in cancer patients<sup>(60)</sup>. Leptin may be considered the antithesis of adiponectin. It is an adipokine that acts centrally to regulate appetite and bodyweight<sup>(61)</sup> and circulating levels are positively associated with adiposity<sup>(62)</sup>. *In vitro* studies confirm promotion of cell proliferation, angiogenesis and matrix metalloproteinase expression in oesophageal and colonic cancer cell lines<sup>(63,64)</sup>. Furthermore, serum levels of leptin are positively correlated with cancer risk<sup>(65)</sup>. Increased production of leptin and decreased production of adiponectin, correlating with amounts of visceral adiposity and the MetSyn, have been associated with cancer promoting pathways including cellular proliferation, angiogenesis and metalloproteinase expression across a wide variety of cancer subtypes<sup>(50,59,63)</sup>.

### Hyperinsulinaemia, insulin resistance and the insulin-like growth factor axis

A core component of the MetSyn is insulin resistance. Nutritionally induced insulin resistance develops as a metabolic adaptation to increased circulating levels of NEFA, which are constantly released from adipose tissue. Increased NEFA levels force liver, muscle and other tissues to shift towards increased storage and oxidation of fats for their energy production. Consequently, insulin secretion rises to compensate for the decreased capacity to handle glucose<sup>(10,66,67)</sup>. Furthermore, there is a decreased

expression of insulin-receptor levels and reduced intracellular insulin signalling in response to insulin receptor binding<sup>(68)</sup> resulting in hyperinsulinaemia and insulin resistance.

The ratio of visceral to subcutaneous fat is a proxy measure of insulin resistance. The rate of lipolysis is higher in visceral adipose tissue than subcutaneous adipose tissue, leading to increased circulation of NEFA<sup>(69)</sup>. Therefore, the risk of developing insulin resistance and hyperinsulinaemia increases with increasing visceral adipose tissue accumulation. Adipokines are believed to be central to the pathogenesis of insulin resistance<sup>(70)</sup>, with high concentrations of TNF- $\alpha$ , IL-6, IL-1 $\beta$ , and low concentrations of adiponectin, having deleterious effects on glucose homeostasis leading to chronic hyperinsulinaemia and insulin resistance<sup>(71)</sup>. A number of epidemiological studies have reported that insulin resistance status, characterised by hyperinsulinaemia, is associated with an increased risk of malignancy, including carcinomas of the breast, prostate and colon<sup>(27,60,72,73)</sup>. Data from the Me-Can study reported that every 1 mmol/l increment in glucose increased risk of incident cancer in men (RR 1.05, 95% CI 1.01, 1.1) and women (RR 1.11, 95% CI 1.05, 1.16) and further increased RR for fatal cancer (men: RR 1.15, 95% CI 1.07, 1.22; women: RR 1.21, 95% CI 1.11, 1.33)<sup>(72)</sup>. Interestingly, studies of type 2 diabetics have found that the risk of colorectal cancer is higher in those treated with insulin<sup>(74)</sup>.

The tumorigenic effects of insulin could be directly mediated by insulin receptors in the pre-neoplastic target cells, or may relate to changes in endogenous hormone metabolism, secondary to hyperinsulinaemia<sup>(23)</sup>. *In vitro* studies demonstrate that insulin increases the neoplastic proliferation of cell lines at both physiological and pharmacological doses, and the insulin receptor is commonly expressed in human neoplasms<sup>(75)</sup>. Whether there are differential downstream signalling effects in normal or transformed epithelial cells compared to insulin-responsive tissues (such as fat, liver and muscle) is the subject of current research, addressing in particular whether receptor activation results in cell survival and proliferation rather than altered energy metabolism<sup>(76)</sup>.

The IGF system may also be relevant to the effects of hyperinsulinaemia in pro-tumour pathways. In the IGF-axis, binding of IGF-1 or IGF-2 to the IGF-1 receptor leads to cell proliferation, differentiation and protection from apoptosis and a dysregulation in this pathway can give rise to malignancy<sup>(77,78)</sup>. Levels of IGF are influenced by circulating insulin levels, with increasing insulin leading to decreased levels of IGF-binding proteins 1 and 2, thus increasing the bioavailability of IGF<sup>(79)</sup>. The IGF-1 axis has been implicated in the progression of breast, pancreatic, colorectal and oesophageal cancer<sup>(80–83)</sup>. Obesity increases levels of bioactive-free IGF-1<sup>(84,85)</sup>, high levels of which are associated with increased risk of malignancy.

In experimental murine models, diet-induced obesity increased both local tumour growth and metastases in wild-type mice compared with lean controls, an effect not seen in liver-specific IGF-1-deficient (LID) mice. In addition, chronic IGF-1 deficiency negates the enhanced expression of inflammatory cytokines and cell adhesion molecules

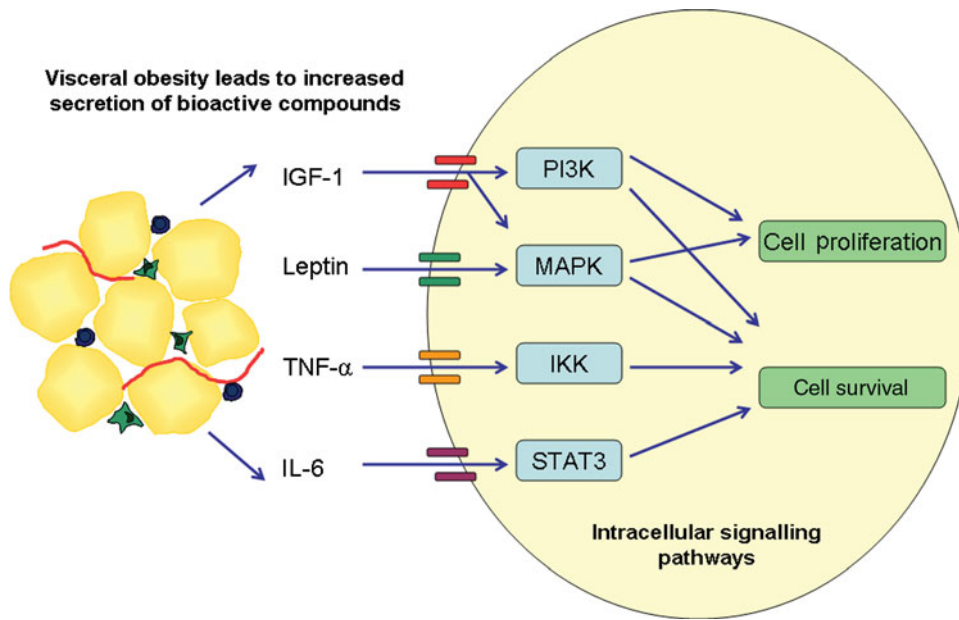
normally up-regulated in obese compared with lean mice<sup>(86–88)</sup>. Hence, it is possible that in obesity IGF-1 can affect tumour development both directly, by stimulating tumour growth and indirectly, by creating a micro-environment that is permissive for tumour growth.

### Cancer cell signalling

Eukaryotic cells coordinate cell growth in line with the availability of nutrients in their environment<sup>(89)</sup>. Obesity as a condition of both systemic insulin resistance and nutrient excess may lead to the activation of intracellular pathways that promote tumour growth and progression. The systemic alterations associated with obesity include a change in inflammatory, sex hormone, insulin and adipokine secretion which may directly influence the tumour micro-environment. Cancers that arise in this environment may go on to develop progressive mutations and epigenetic alterations which are influenced by this obese milieu. The concept of oncogene addiction describes the apparent dependency of some tumours on one or a few genes to maintain the malignant phenotype<sup>(90)</sup>. Clinical evidence of this is cited as the ability of therapies targeting specific genes or pathways to inhibit cancer cell growth or improve survival rates<sup>(91)</sup>. Whole genome arrays have demonstrated that breast cancer may be subdivided into a number of types based on the genes over-expressed in certain subtypes and that targeting these predominant pathways provides avenues for chemotherapy treatment for each subtype<sup>(92–94)</sup>. Molecular characterisation of tumours has been exploited to develop assays to predict subgroups of patients with poorer prognosis who may benefit from adjuvant therapy<sup>(95)</sup>.

Cancers that develop within an obese environment may become selectively altered to signal via specific pathways. For examples, in patients with non-small cell lung cancer, only a subset (approximately 10–20%) respond to the epidermal growth factor receptor targeted therapy gefitinib, and these patients often have an activating mutation of epidermal growth factor receptor<sup>(96)</sup>. Patients with activating mutations are more likely to have adenocarcinomas, and to be female, non-smokers and Japanese<sup>(97)</sup>. Similarly, obesity-related cancers may have a specific set of targets (malfunctioning molecules or pathways), which may be exploitable in clinical practice. Certainly, a number of the putatively dysregulated adipokines and growth factors in obesity may signal via the same intracellular signalling pathways. Candidate pathways include the phosphoinositide 3-kinase (PI3K), mitogen-activated protein kinase and signal transducer and activator of transcription 3 pathways. Activation of these pathways leads to multiple downstream effects that underpin cancer progression and metastasis<sup>(98–100)</sup>. Importantly, inhibitors of these pathways are under development at present in order to provide new therapeutic avenues<sup>(101–103)</sup>.

Is there any evidence that cancers which develop within the obese milieu are 'addicted to' or preferentially signal via these pathways? Whole genome analysis of breast cancer tumours divided according to BMI demonstrate that an obesity-associated gene signature pattern is associated



**Fig. 2.** (Colour online) Visceral adiposity effects cancer cell signalling pathways. Increased visceral adipose tissue leads to increased circulation of a number of bioactive compounds such as IL-6, TNF- $\alpha$ , leptin and insulin-like growth factor-1 (IGF-1). Binding of these compounds to their respective receptors on tumour cells leads to the activation of cell signalling pathways including the phosphatidylinositol 3-kinase (PI3K), mitogen-activated-protein-kinase (MAPK), signal transducer and activator of transcription 3 (STAT3) and I $\kappa$ B kinase (IKK) pathways. The cascade of downstream signalling leads to increased cell survival and proliferation thus promoting tumour progression.

with a shorter time to metastasis and is associated with an IGF signalling signature in multiple publically available breast cancer genome arrays<sup>(104)</sup>. Most data come from murine models. PI3K activity (measured by phosphorylated protein kinase B and mammalian target of rapamycin protein levels) is increased in diet-induced obesity in mice and is associated with an increased level of circulating IGF-1 compared with controls<sup>(105)</sup>. Mice fed a high-energy diet have twice the volume of tumours 17 d after colon cancer cell injection versus controls. PI3K pathway activity was demonstrated by increased phosphorylated protein kinase B protein levels. The tumour growth effect was abrogated by metformin treatment, which led to decreased phosphorylated protein kinase B levels<sup>(106)</sup>. In a mouse model of obesity-related skin cancer, obese mice had higher PI3K activity after UV exposure than lean mice<sup>(107)</sup>. Activity of mitogen-activated protein kinase phosphorylation and NF- $\kappa$ B signalling were also higher following UVB irradiation in the obese mice<sup>(108)</sup>. In breast and colorectal cancer, over-expression of the leptin receptor has been identified. As leptin has mitogenic and anti-apoptotic effects on cancer cells lines it is postulated that it acts through mitogen-activated protein kinase and PI3K pathways. This hypothesis is strengthened by the observation that inhibition of these pathways attenuates the growth promoting effects of leptin<sup>(109)</sup>. In an obesity-associated hepatoma model, IL-6 and TNF- $\alpha$  induce the development of cancer via activation of signal transducer and activator of transcription 3 pathway<sup>(110)</sup>. Excess energy balance associated with the obese state may also influence tumour growth. Mouse tumour xenografts have decreased incidence and slower growth in mice that are fed an

energy-restricted diet. Tumours that are resistant to dietary restriction have constitutive activation of the PI3K pathway<sup>(111)</sup>. The proposed cell signalling pathways that are altered in the obese state and associated with tumorigenesis are illustrated in Fig. 2.

### Visceral obesity, inflammation and cancer

As early as the nineteenth century it was perceived that cancer is linked to inflammation and an inflammatory component is present in the microenvironment of most neoplastic tissues. Key features of cancer-related inflammation include the infiltration of immune cells, the presence of cytokines and chemokines, including TNF- $\alpha$ , IL-1 $\beta$ , IL-6 and chemokines such as monocyte chemoattractant protein-1 and CXCL8<sup>(112)</sup>. Thus, cancer-related inflammation is a key component of the tumorigenic process and is increasingly referred to as the seventh hallmark of cancer.

NF- $\kappa$ B, first described in 1986 by Ranjan Sen and David Baltimore, is a central coordinator for multiple inflammatory and anti-inflammatory signalling pathways and has become one of the most investigated transcription factors<sup>(113)</sup>. Activation of NF- $\kappa$ B has been shown to control a range of cellular processes in cancer including inflammation, proliferation, angiogenesis, metastasis, chemoresistance and radioresistance<sup>(114,115)</sup>. Obesity has been shown to increase the systemic activation of NF- $\kappa$ B<sup>(116)</sup>. It is constitutively active in a wide range of human tumour cells, and its suppression has been shown to inhibit their growth, leading to the 'NF- $\kappa$ B addiction' theory in tumour cells<sup>(114)</sup>.

The omentum is one of the largest components of the visceral adipose depot in human subjects and has been the subject of research for more than 3600 years<sup>(117)</sup>. The word 'omentum' has its origins in the practice of Egyptian priests examining the abdominal viscera during embalming the body. They would tell fortunes by it and give good or bad 'omens' for the deceased<sup>(117)</sup>. Adipose tissue, including the omentum, is thought to be a primitive immune organ, but unlike other adipose tissue depots the omentum has many unique functions<sup>(118)</sup>. It migrates to and surrounds areas of inflammation and infection within the peritoneal cavity and is a rich source of pro-angiogenic and inflammatory mediators<sup>(119)</sup>. Dense lymphoreticular structures within the human omentum, termed milky spots contain a range of innate and adaptive immune cells, namely macrophages, mast cells, T- and B-cells<sup>(120)</sup>. More recent studies have also demonstrated the presence of natural killer and invariant natural killer T-cell populations within the human omentum, which are altered by obesity and cancer<sup>(121)</sup>. These innate lymphocytes are capable of killing virally infected or tumour cells in a non-antigen-specific manner. In addition to an important role in immunosurveillance of the peritoneal cavity, another key function of the human omentum is to support B-cell function. The omentum collects antigens and cells from the peritoneal cavity and supports T-dependent B-cell responses, including isotype switching, somatic hypermutation and limited affinity maturation, despite the lack of identifiable follicular dendritic cells<sup>(122)</sup>.

Macrophages are the most abundant leukocyte population in adipose tissue<sup>(123)</sup>. Infiltration of inflammatory macrophages into adipose tissues correlates with increasing obesity levels and triggers insulin resistance and fuels inflammation. A significant number of macrophages are also present in lean, metabolically healthy human subjects suggesting that not all adipose tissue macrophages are programmed to be pro-inflammatory. Current evidence indicates that macrophages undergo a functional conversion to promote inflammation during obesity<sup>(124)</sup>. In visceral fat, obesity alters the state of adipose tissue macrophages from an anti-inflammatory M2 phenotype to a pro-inflammatory M1 state<sup>(123)</sup>, further potentiating metabolic dysfunction, insulin resistance and inflammation in obese individuals.

The majority of studies so far evaluating the immune cell properties of visceral adipose tissue have predominantly focused on macrophages. T-cells, however, may be key regulators of adipose tissue inflammation, and a recent study in a murine model revealed a significant increase in CD8<sup>+</sup> T-cell infiltration into expanding visceral adipose tissue, which preceded macrophage infiltration<sup>(125)</sup>. These T-cells were found to be responsible for both the establishment and maintenance of adipose tissue inflammation in obesity<sup>(125)</sup>. We have recently shown that the human omentum is a particularly rich source of CD8<sup>+</sup> T-cells in patients with obesity-associated oesophageal adenocarcinoma. A high percentage of CD4<sup>+</sup> and CD8<sup>+</sup> omental T-cells were activated with an inflammatory T-helper 1 phenotype (CD69<sup>+</sup>CD45RO<sup>+</sup>IFN- $\gamma$ <sup>+</sup>)<sup>(126)</sup>. Interestingly, we also demonstrated that significantly more interferon- $\gamma$ <sup>+</sup> T-cells were present in the omentum of cancer patients

compared with non-cancer controls<sup>(126)</sup>. The role of T-cells in adipose tissue inflammation and cancer has not received a lot of attention so far but offers potential immunotherapeutic targets for obesity-associated morbidities.

## Conclusion

Adipose tissue is not simply an energy store but acts in a co-ordinated fashion to exert systemic hormonal and metabolic effects. There is sound epidemiological evidence relating excess adiposity and associated dysmetabolism to a broad range of cancer subtypes and emerging clinical research continues to bolster this observation. The recognition of visceral obesity as a major contributing factor to the development of cancer mandates further research into pathophysiological mechanisms. Much work remains to elucidate the independent and synergistic effects of inflammation, insulin resistance, the IGF-axis and adipokines on carcinogenesis in obesity. Clearly, the association between obesity and cancer is an important area to target in public health policy and one that will assume increasing importance over the coming years.

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## References

1. World Health Organisation (2000) *Obesity: Preventing and Managing the Global Epidemic. Report of a WHO Consultation*. World Health Organisation Technical Report Series 894, pp. 1–253. Geneva: WHO.
2. Ogden CL, Carroll MD, Curtin LR *et al.* (2006) Prevalence of overweight and obesity in the united states, 1999–2004. *JAMA* **295**, 1549–1555.
3. National Health Service Information Centre (2009) Health survey for England–2008: Physical activity and fitness. <http://www.ic.nhs.uk/statistics-and-data-collections/health-and-lifestyles-related-surveys/health-survey-for-england/health-survey-for-england-2008-physical-activity-and-fitness> (accessed August 2011).
4. McCarthy SN, Gibney MJ & Flynn A (2002) Overweight, obesity and physical activity levels in Irish adults: Evidence from the North/South Ireland food consumption survey. *Proc Nutr Soc* **61**, 3–7.
5. Jackson-Leach R & Lobstein T (2006) Estimated burden of paediatric obesity and co-morbidities in Europe. Part 1. The

- increase in the prevalence of child obesity in Europe is itself increasing. *Int J Pediatr Obes* **1**, 26–32.
6. Serdula MK, Ivery D, Coates RJ *et al.* (1993) Do obese children become obese adults? A review of the literature. *Prev Med* **22**, 167–177.
  7. Klein S, Warden T & Superman HJ (2002) AGA technical review on obesity. *Gastroenterology* **123**, 882–932.
  8. René an AG, Tyson M, Egger M *et al.* (2008) Body-mass index and incidence of cancer: A systematic review and meta-analysis of prospective observational studies. *Lancet* **371**, 569–578.
  9. World Cancer Research Fund, American Institute for Cancer Research (2007) *Food, Nutrition, Physical Activity and the Prevention of Cancer: Global Perspective*. Washington, DC: American Institute for Cancer Research.
  10. Calle EE, Rodriguez C, Walker-Thurmond K *et al.* (2003) Overweight, obesity, and mortality from cancer in a prospectively studied cohort of U.S. Adults. *N Engl J Med* **348**, 1625–1638.
  11. Dirx MJM, Zeegers MPA, Dagnelie PC *et al.* (2003) Energy restriction and the risk of spontaneous mammary tumors in mice: A meta-analysis. *Int J Cancer* **106**, 766–770.
  12. Mai V, Colbert LH, Berrigan D *et al.* (2003) Calorie restriction and diet composition modulate spontaneous intestinal tumorigenesis in Apc(Min) mice through different mechanisms. *Cancer Res* **63**, 1752–1755.
  13. Sjöström L, Gummesson A, Sjöström CD *et al.* (2009) Effects of bariatric surgery on cancer incidence in obese patients in Sweden (Swedish obese subjects study): A prospective, controlled intervention trial. *Lancet Oncol* **10**, 653–662.
  14. Vague J (1956) The degree of masculine differentiation of obesities: A factor determining predisposition to diabetes, atherosclerosis, gout, and uric calculous disease. *Obes Res* **4**, 204–212.
  15. Vague J (1947) La différenciation sexuelle, facteur déterminant des formes de l'obésité. *Presse Med* **53**, 339–340.
  16. Fox CS, Massaro JM, Hoffmann U *et al.* (2007) Abdominal visceral and subcutaneous adipose tissue compartments. *Circulation* **116**, 39–48.
  17. Galic S, Oakhill JS & Steinberg GR (2010) Adipose tissue as an endocrine organ. *Mol Cell Endocrinol* **316**, 129–139.
  18. Alberti KGMM, Zimmet P & Shaw J (2006) Metabolic syndrome – a new world-wide definition. A Consensus Statement from the International Diabetes Federation. *Diabet Med* **23**, 469–480.
  19. Fujioka S, Matsuzawa Y, Tokunaga K *et al.* (1987) Contribution of intra-abdominal fat accumulation to the impairment of glucose and lipid metabolism in human obesity. *Metabolism* **36**, 54–59.
  20. Nedungadi TP & Clegg DJ (2009) Sexual dimorphism in body fat distribution and risk for cardiovascular diseases. *J Cardiovasc Transl Res* **2**, 321–327.
  21. Schapira DV, Clark RA, Wolff PA *et al.* (1994) Visceral obesity and breast cancer risk. *Cancer* **74**, 632–639.
  22. Beddy P, Howard J, McMahon C *et al.* (2010) Association of visceral adiposity with oesophageal and junctional adenocarcinomas. *Br J Surg* **97**, 1028–1034.
  23. Schoen RE, Tangen CM, Kuller LH *et al.* (1999) Increased blood glucose and insulin, body size, and incident colorectal cancer. *J Natl Cancer Inst* **91**, 1147–1154.
  24. Yamaji T, Iwasaki M, Sasazuki S *et al.* (2009) Visceral fat volume and the prevalence of colorectal adenoma. *Am J Epidemiol* **170**, 1502–1511.
  25. Nam SY, Kim BC, Han KS *et al.* (2010) Abdominal visceral adipose tissue predicts risk of colorectal adenoma in both sexes. *Clin Gastroenterol Hepatol* **8**, 443–450.
  26. Cowey S & Hardy RW (2006) The metabolic syndrome: A high-risk state for cancer? *Am J Pathol* **169**, 1505–1522.
  27. Giovannucci E (2007) Metabolic syndrome, hyperinsulinemia, and colon cancer: A review. *Am J Clin Nutr* **86**, 836S–842S.
  28. Colangelo LA, Gapstur SM, Gann PH *et al.* (2002) Colorectal cancer mortality and factors related to the insulin resistance syndrome. *Cancer Epidemiol Biomarkers Prev* **11**, 385–391.
  29. Bowers K, Albanes D, Limburg P *et al.* (2006) A prospective study of anthropometric and clinical measurements associated with insulin resistance syndrome and colorectal cancer in male smokers. *Am J Epidemiol* **164**, 652–664.
  30. Stocks T, Borena W, Strohmaier S *et al.* (2010) Cohort profile: The metabolic syndrome and cancer project (Me-Can). *Int J Epidemiol* **39**, 660–667.
  31. Stocks T, Lukanova A, Bjørge T *et al.* (2010) Metabolic factors and the risk of colorectal cancer in 580,000 men and women in the metabolic syndrome and cancer project (Me-Can). *Cancer* **117**, 2398–2407.
  32. Bjørge T, Stocks T, Lukanova A *et al.* (2010) Metabolic syndrome and endometrial carcinoma. *Am J Epidemiol* **171**, 892–902.
  33. Häggström C, Stocks T, Rapp K *et al.* (2011) Metabolic syndrome and risk of bladder cancer: Prospective cohort study in the metabolic syndrome and cancer project (Me-Can). *Int J Cancer* **128**, 1890–1898.
  34. Johansen D, Stocks T, Jonsson HK *et al.* (2010) Metabolic factors and the risk of pancreatic cancer: A prospective analysis of almost 580,000 men and women in the metabolic syndrome and cancer project. *Cancer Epidemiol Biomarkers Prev* **19**, 2307–2317.
  35. Russo A, Autelitano M & Bisanti L (2008) Metabolic syndrome and cancer risk. *Eur J Cancer* **44**, 293–297.
  36. Kang HW, Kim D, Kim HJ *et al.* (2009) Visceral obesity and insulin resistance as risk factors for colorectal adenoma: A cross-sectional, case-control study. *Am J Gastroenterol* **105**, 178–187.
  37. Siegel EM, Ulrich CM, Poole EM *et al.* (2010) The effects of obesity and obesity-related conditions on colorectal cancer prognosis. *Cancer Control* **17**, 52–57.
  38. Healy L, Howard J, Ryan A *et al.* (2011) Metabolic syndrome and leptin are associated with adverse pathological features in male colorectal cancer patients. *Colorectal Dis* **22**, 281–288.
  39. Healy LA, Ryan AM, Carroll P *et al.* (2010) Metabolic syndrome, central obesity and insulin resistance are associated with adverse pathological features in postmenopausal breast cancer. *Clin Oncol (R Coll Radiol)* **22**, 281–288.
  40. Shen Z, Wang S, Ye Y *et al.* (2010) Clinical study on the correlation between metabolic syndrome and colorectal carcinoma. *ANZ J Surg* **80**, 331–336.
  41. Moon H-G, Ju Y-T, Jeong C-Y *et al.* (2008) Visceral obesity may affect oncologic outcome in patients with colorectal cancer. *Ann Surg Oncol* **15**, 1918–1922.
  42. Jankowski JA, Wright NA, Meltzer SJ *et al.* (1999) Molecular evolution of the metaplasia–dysplasia–adenocarcinoma sequence in the esophagus. *Am J Pathol* **154**, 965–973.
  43. National Cancer Registry of Ireland (2009) <http://www.ncri.ie/data> (accessed August 2011).
  44. Bollschweiler E, Wolfgarten E, Gutschow C *et al.* (2001) Demographic variations in the rising incidence of esophageal adenocarcinoma in white males. *Cancer* **92**, 549–555.
  45. Calle EE & Kaaks R (2004) Overweight, obesity and cancer: Epidemiological evidence and proposed mechanisms. *Nat Rev Cancer* **4**, 579–591.

46. Friedenberg FK, Xanthopoulos M, Foster GD *et al.* (2008) The association between gastroesophageal reflux disease and obesity. *Am J Gastroenterol* **103**, 2111–2122.
47. Lagergren J, Bergstrom R & Nyren O (1999) Association between body mass and adenocarcinoma of the esophagus and gastric cardia. *Ann Intern Med* **130**, 883–890.
48. Ryan AM, Rowley SP, Fitzgerald AP *et al.* (2006) Adenocarcinoma of the oesophagus and gastric cardia: Male preponderance in association with obesity. *Eur J Cancer* **42**, 1151–1158.
49. Ryan AM, Healy LA, Power DG *et al.* (2008) Barrett esophagus: Prevalence of central adiposity, metabolic syndrome, and a proinflammatory state. *Ann Surg* **247**, 909–915.
50. Howard JM, Beddy P, Ennis D *et al.* (2010) Associations between leptin and adiponectin receptor upregulation, visceral obesity and tumour stage in oesophageal and junctional adenocarcinoma. *Br J Surg* **97**, 1020–1027.
51. Elliott BE, Tam SP, Dexter D *et al.* (1992) Capacity of adipose tissue to promote growth and metastasis of a murine mammary carcinoma: Effect of estrogen and progesterone. *Int J Cancer* **51**, 416–424.
52. Celis JE, Moreira JM, Cabezon T *et al.* (2005) Identification of extracellular and intracellular signalling components of the mammary adipose tissue and its interstitial fluid in high risk breast cancer patients: Toward dissecting the molecular circuitry of epithelial-adipocyte stromal cell interactions. *Mol Cell Proteomics* **4**, 492–522.
53. Cook KS, Min HY, Johnson D *et al.* (1987) Adipsin: A circulating serine protease homolog secreted by adipose tissue and sciatic nerve. *Science* **237**, 402–405.
54. Flier JS, Cook KS, Usher P *et al.* (1987) Severely impaired adipsin expression in genetic and acquired obesity. *Science* **237**, 405–408.
55. Zhang Y, Proenca R, Maffei M *et al.* (1994) Positional cloning of the mouse obese gene and its human homologue. *Nature* **372**, 425–432.
56. Scherer PE, Williams S, Fogliano M *et al.* (1995) A novel serum protein similar to C1q, produced exclusively in adipocytes. *J Biol Chem* **270**, 26746–26749.
57. Ouchi N, Parker JL, Lugus JJ *et al.* (2011) Adipokines in inflammation and metabolic disease. *Nat Rev Immunol* **11**, 85–97.
58. Harvey AE, Lashinger LM & Hursting SD (2011) The growing challenge of obesity and cancer: An inflammatory issue. *Ann NY Acad Sci* **1229**, 45–52.
59. Kadowaki T & Yamauchi T (2005) Adiponectin and adiponectin receptors. *Endocr Rev* **26**, 439–451.
60. Rose DP, Komninou D & Stephenson GD (2004) Obesity, adipocytokines, and insulin resistance in breast cancer. *Obes Rev* **5**, 153–165.
61. Cummings DE & Foster KE (2003) Ghrelin–leptin tango in body-weight regulation. *Gastroenterology* **124**, 1532–1535.
62. Pär S, Annkatrin L, Carine B *et al.* (2004) Obesity and colon cancer: Does leptin provide a link? *Int J Cancer* **109**, 149–152.
63. Somasundar P, McFadden DW, Hileman SM *et al.* (2004) Leptin is a growth factor in cancer. *J Surg Res* **116**, 337–349.
64. Howard JM, Pidgeon GP & Reynolds JV (2010) Leptin and gastro-intestinal malignancies. *Obes Rev* **11**, 863–874.
65. Garofalo C & Surmacz E (2006) Leptin and cancer. *J Cell Physiol* **207**, 12–22.
66. Bergman RN & Ader M (2000) Free fatty acids and pathogenesis of type 2 diabetes mellitus. *Trends Endocrinol Metab* **11**, 351–356.
67. Ebeling P & Koivisto V (1994) Non-esterified fatty acids regulate lipid and glucose oxidation and glycogen synthesis in healthy man. *Diabetologia* **37**, 202–209.
68. Moller DE & Flier JS (1991) Insulin resistance – mechanisms, syndromes, and implications. *N Engl J Med* **325**, 938–948.
69. Wajchenberg BL (2000) Subcutaneous and visceral adipose tissue: Their relation to the metabolic syndrome. *Endocr Rev* **21**, 697–738.
70. Fasshauer M & Paschke R (2003) Regulation of adipocytokines and insulin resistance. *Diabetologia* **46**, 1594–1603.
71. Greenberg AS & McDaniel ML (2002) Identifying the links between obesity, insulin resistance and beta-cell function: Potential role of adipocyte-derived cytokines in the pathogenesis of type 2 diabetes. *Eur J Clin Invest* **32**, Suppl. 3, 24–34.
72. Stocks T, Rapp K, Bjørge T *et al.* (2009) Blood glucose and risk of incident and fatal cancer in the metabolic syndrome and cancer project (Me-Can): Analysis of six prospective cohorts. *PLoS Med* **6**, e1000201.
73. Hsing AW, Gao Y-T, Chua S *et al.* (2003) Insulin resistance and prostate cancer risk. *J Natl Cancer Inst* **95**, 67–71.
74. Yang Y-X, Hennessy S & Lewis JD (2004) Insulin therapy and colorectal cancer risk among type 2 diabetes mellitus patients. *Gastroenterology* **127**, 1044–1050.
75. Osborne CK, Bolan G, Monaco ME *et al.* (1976) Hormone responsive human breast cancer in long-term tissue culture: Effect of insulin. *Proc Natl Acad Sci USA* **73**, 4536–4540.
76. Pollak M (2008) Insulin and insulin-like growth factor signalling in neoplasia. *Nat Rev Cancer* **8**, 915–928.
77. Frasca F, Pandini G, Sciacca L *et al.* (2008) The role of insulin receptors and IGF-1 receptors in cancer and other diseases. *Arch Physiol Biochem* **114**, 23–37.
78. Samani AA, Yakar S, LeRoith D *et al.* (2007) The role of the IGF system in cancer growth and metastasis: Overview and recent insights. *Endocr Rev* **28**, 20–47.
79. Jones JI & Clemmons DR (1995) Insulin-like growth factors and their binding proteins: Biological actions. *Endocr Rev* **16**, 3–34.
80. Coussens LM & Werb Z (2002) Inflammation and cancer. *Nature* **420**, 860–867.
81. Chan JM, Stampfer MJ, Giovannucci E *et al.* (1998) Plasma insulin-like growth factor-1 and prostate cancer risk: A prospective study. *Science* **279**, 563–566.
82. Ma J, Pollak MN, Giovannucci E *et al.* (1999) Prospective study of colorectal cancer risk in men and plasma levels of insulin-like growth factor (IGF)-1 and IGF-binding protein-3. *J. Natl. Cancer Inst.* **91**, 620–625.
83. Renehan AG, Frystyk J & Flyvbjerg A (2006) Obesity and cancer risk: The role of the insulin-IGF axis. *Trends Endocrinol Metab* **17**, 328–336.
84. Frystyk J, Vestbo E, Skjaerbaek C *et al.* (1995) Free insulin-like growth factors in human obesity. *Metabolism* **44**, 37–44.
85. Nam SY, Lee EJ, Kim KR *et al.* (1997) Effect of obesity on total and free insulin-like growth factor (IGF)-1, and their relationship to IGF-binding protein (BP)-1, IGFBP-2, IGFBP-3, insulin, and growth hormone. *Int J Obes (Lond)* **21**, 355.
86. Wu Y, Brodt P, Sun H *et al.* (2010) Insulin-like growth factor-1 regulates the liver microenvironment in obese mice and promotes liver metastasis. *Cancer Res* **70**, 57–67.
87. Lashinger LM, Malone LM, McArthur MJ *et al.* (2011) Genetic reduction of insulin-like growth factor-1 mimics the anticancer effects of calorie restriction on cyclooxygenase-2-driven pancreatic neoplasia. *Cancer Prev Res (Phila)* **4**, 1030–1040.



88. Olivo-Marston SE, Hursting SD, Lavigne J *et al.* (2009) Genetic reduction of circulating insulin-like growth factor-1 inhibits azoxymethane-induced colon tumorigenesis in mice. *Mol Carcinog* **48**, 1071–1076.
89. Shaw RJ & Cantley LC (2006) Ras, PI(3)K and MTOR signalling controls tumour cell growth. *Nature* **441**, 424–430.
90. Weinstein IB (2002) Addiction to oncogenes – the achilles heel of cancer. *Science* **297**, 63–64.
91. Weinstein IB & Joe AK (2006) Mechanisms of disease: Oncogene addiction – a rationale for molecular targeting in cancer therapy. *Nat Clin Prac Oncol* **3**, 448–457.
92. Perou CM, Sorlie T, Eisen MB *et al.* (2000) Molecular portraits of human breast tumours. *Nature* **406**, 747–752.
93. Sørlie T, Tibshirani R, Parker J *et al.* (2003) Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proc Natl Acad Sci USA* **100**, 8418–8423.
94. Sawyers C (2004) Targeted cancer therapy. *Nature* **432**, 294–297.
95. Paik S, Shak S, Tang G *et al.* (2004) A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. *New Engl J Med* **351**, 2817–2826.
96. Lynch TJ, Bell DW, Sordella R *et al.* (2004) Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N Engl J Med* **350**, 2129–2139.
97. Taron M, Ichinose Y, Rosell R *et al.* (2005) Activating mutations in the tyrosine kinase domain of the epidermal growth factor receptor are associated with improved survival in gefitinib-treated chemorefractory lung adenocarcinomas. *Clin Cancer Res* **11**, 5878–5885.
98. Huang P, Han J & Hui L (2010) MAPK signalling in inflammation-associated cancer development. *Protein Cell* **1**, 218–226.
99. Yu H, Pardoll D & Jove R (2009) STATs in cancer inflammation and immunity: A leading role for STAT3. *Nat Rev Cancer* **9**, 798–809.
100. Aggarwal BB, Kunnumakkara AB, Harikumar KB *et al.* (2009) Signal transducer and activator of transcription-3, inflammation, and cancer. *Ann NY Acad Sci* **1171**, 59–76.
101. Sebolt-Leopold JS & Herrera R (2004) Targeting the mitogen-activated protein kinase cascade to treat cancer. *Nat Rev Cancer* **4**, 937–947.
102. Liu P, Cheng H, Roberts TM *et al.* (2009) Targeting the phosphoinositide 3-kinase pathway in cancer. *Nat Rev Drug Discov* **8**, 627–644.
103. Jing N & Twardy DJ (2005) Targeting STAT3 in cancer therapy. *Anticancer Drugs* **16**, 601–607.
104. Creighton CJ, Sada YH, Zhang Y *et al.* (2011) A gene transcription signature of obesity in breast cancer. *Breast Cancer Res Treat* (Epublication ahead of print version).
105. Moore T, Beltran L, Carbajal S *et al.* (2008) Dietary energy balance modulates signalling through the Akt/mammalian target of rapamycin pathways in multiple epithelial tissues. *Cancer Prev Res (Phila)* **1**, 65–76.
106. Algire C, Amrein L, Zakikhani M *et al.* (2010) Metformin blocks the stimulative effect of a high-energy diet on colon carcinoma growth *in vivo* and is associated with reduced expression of fatty acid synthase. *Endocr Relat Cancer* **17**, 351–360.
107. Sharma SD & Katiyar SK (2010) Leptin deficiency-induced obesity exacerbates ultraviolet b radiation-induced cyclooxygenase-2 expression and cell survival signals in ultraviolet b-irradiated mouse skin. *Toxicol Appl Pharmacol* **244**, 328–335.
108. Katiyar SK & Meeran SM (2007) Obesity increases the risk of UV radiation-induced oxidative stress and activation of MAPK and NF-kappaB signalling. *Free Radic Biol Med* **42**, 299–310.
109. van Kruijsdijk RCM, van der Wall E & Visseren FLJ (2009) Obesity and cancer: The role of dysfunctional adipose tissue. *Cancer Epidemiol Biomarkers Prev* **18**, 2569–2578.
110. Park EJ, Lee JH, Yu G-Y *et al.* (2010) Dietary and genetic obesity promote liver inflammation and tumorigenesis by enhancing IL-6 and TNF expression. *Cell* **140**, 197–208.
111. Kalaany NY & Sabatini DM (2009) Tumours with PI3K activation are resistant to dietary restriction. *Nature* **458**, 725–731.
112. Colotta F, Allavena P, Sica A *et al.* (2009) Cancer-related inflammation, the seventh hallmark of cancer: Links to genetic instability. *Carcinogenesis* **30**, 1073–1081.
113. Sen R & Baltimore D (1986) Multiple nuclear factors interact with the immunoglobulin enhancer sequences. *Cell* **46**, 705–716.
114. Chaturvedi MM, Sung B, Yadav VR *et al.* (2011) NF-κB addiction and its role in cancer: ‘one size does not fit all’. *Oncogene* **30**, 1615–1630.
115. Ben-Neriah Y & Karin M (2011) Inflammation meets cancer, with NF-κB as the matchmaker. *Nat Immunol* **12**, 715–723.
116. Cai D, Yuan M, Frantz DF *et al.* (2005) Local and systemic insulin resistance resulting from hepatic activation of IKK-β and NF-κB. *Nat Med* **11**, 183–190.
117. Walker FC (1963) The protective function of the greater omentum. *Ann R Coll Surg Engl* **33**, 282–306.
118. Caspar-Bauguil S, Cousin B, Galinier A *et al.* (2005) Adipose tissues as an ancestral immune organ: Site-specific change in obesity. *FEBS Lett* **579**, 3487–3492.
119. Casten DF & Alday ES (1971) Omental transfer for revascularization of the extremities. *Surg Gynecol Obstet* **132**, 301–304.
120. Platell C, Cooper D, Papadimitriou JM *et al.* (2000) The omentum. *World J Gastroenterol* **6**, 169–176.
121. Lynch L, O’Shea D, Winter DC *et al.* (2009) Invariant NKT cells and CD1d+ cells amass in human omentum and are depleted in patients with cancer and obesity. *Eur J Immunol* **39**, 1893–1901.
122. Rangel-Moreno J, Moyron-Quiroz JE, Carragher DM *et al.* (2009) Omental milky spots develop in the absence of lymphoid tissue-inducer cells and support B and T cell responses to peritoneal antigens. *Immunity* **30**, 731–743.
123. Morris DL, Singer K & Lumeng CN (2011) Adipose tissue macrophages: Phenotypic plasticity and diversity in lean and obese states. *Curr Opin Clin Nutr Metab Care* **14**, 341–346.
124. Lumeng CN, Bodzin JL & Saltiel AR (2007) Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *J Clin Invest* **117**, 175–184.
125. Nishimura S, Manabe I, Nagasaki M *et al.* (2009) CD8+ effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nat Med* **15**, 914–920.
126. Lysaght J, Allott EH, Donohoe CL *et al.* (2011) T lymphocyte activation in visceral adipose tissue of patients with oesophageal adenocarcinoma. *Br J Surg* **98**, 964–974.