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## When Tumor Suppressor TGF $\beta$ Meets the HER2 (ERBB2) Oncogene

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### Abstract

Despite its tumor suppressive role in normal mammary epithelial cells, TGF $\beta$  has been reported to promote the migration, invasion and survival in breast cancer cells overexpressing the HER2 (ERBB2; neu) oncogene, and to accelerate the metastasis of neu-induced mammary tumors in mice. A clearer understanding of the molecular mechanisms underlying the crosstalk between TGF $\beta$  and HER2 has started to emerge. In recent studies reviewed here, the synergistic effect of TGF $\beta$  and HER2 on tumor progression has been shown to likely be a combined result of two distinct features: (1) loss of TGF $\beta$ 's tumor suppressive effect through functional alterations in the anti-mitogenic effect of Smad-mediated transcription, and (2) gain of pro-survival and pro-migratory function through HER2-dependent mechanisms. In HER2-overexpressing breast cancer, this crosstalk results in increased cancer cell proliferation, survival and invasion, accelerated metastasis in animal models, and resistance to chemotherapy and HER2-targeted therapy. Thus, the transformed cellular context imparted by constitutively active HER2 signaling, as a consequence of HER2 gene amplification or overexpression, aborts the tumor suppressive role of TGF $\beta$  and facilitated the oncogenic role of this pathway. In turn, TGF $\beta$  potentiates oncogenic HER2 signaling by inducing shedding of the ERBB ligands and clustering of HER2 with integrins. Here we discuss recent studies examining Smad-dependent and -independent mechanisms of crosstalk between TGF $\beta$  and HER2. Therefore, blockade of TGF $\beta$ :HER2 crosstalk may suppress breast cancer progression and metastasis, and enhance the efficiency of conventional therapies in patients with HER2-overexpressing breast cancer.

### Keywords

TGF $\beta$ ; HER2 (ERBB2); Breast cancer; Drug resistance

## Introduction

The cellular phenotypes regulated by transforming growth factor  $\beta$  (TGF $\beta$ ) signaling are highly context-dependent, which underly the different and often opposite TGF $\beta$  functions in cancerous and normal cells. TGF- $\beta$  family members signal through the heteromeric complex of trans-membrane serine/threonine kinases, the type I and type II receptors (T $\beta$ RI and T $\beta$ RII), which subsequently phosphorylate receptor-regulated Smads (R-Smads). R-Smads, usually together with a common mediator Smad, Smad4, then translocate to the nucleus where they regulate gene transcription via binding to the promoter of target genes [1]. Non-Smad pathways, including the phosphatidylinositol-3 kinase (PI3K), extracellular signal-regulated kinase (ERK, MAPK), c-Jun NH<sub>2</sub>-terminal kinase (JNK), p38MAPK, and Rho GTPases, have also been implicated in TGF $\beta$  action [2, 3]. Studies by our group and others indicate that one of the contextual factors for TGF $\beta$  function is HER2 (ERBB2; neu, the rat/mouse homologue of HER2), an oncogene frequently activated by gene amplification or overexpression in human breast cancer. In fact, TGF $\beta$  plays a tumor suppressive role in normal epithelia by inhibiting cell proliferation and inducing apoptosis, but accelerates progression of established cancers by autocrine and paracrine mechanisms [2, 4, 5]. The role of TGF $\beta$  in cancer appears to result from two actions: (1) loss of its tumor suppressive effects; and (2) gain of function as a cancer-promoting agent that synergizes with transforming oncogenes. In this review, we discuss recent findings from our group and others on the mechanisms through which HER2, as an example of oncogene-mediated alterations of cellular context, redirects TGF $\beta$ 's function to facilitate cancer progression.

### The HER2 (ERBB2) Oncogene and Breast Cancer

*HER2* gene amplification or overexpression of its product, the receptor tyrosine kinase (RTK) HER2, occurs in approximately 25% of human breast cancers, where it is associated with drug resistance, metastatic behavior, and overall poor patient outcome [6, 7]. HER2 is a member of the ERBB (Erythroblastic Leukemia Viral Oncogene Homolog) receptor family, which also includes the epidermal growth factor receptor (EGFR, ERBB1), HER3 (ERBB3), and HER4 (ERBB4). Ligand binding to the ectodomains of EGFR, ERBB3, and ERBB4 results in the formation of catalytically active homo- and heterodimers to which HER2 is recruited as a preferred partner [8]. Although HER2 does not bind any ERBB ligand directly, its catalytic activity can potently amplify signaling by ERBB-containing heterodimers via increasing ligand binding affinity and/or receptor recycling and stability [9–12]. Activation of the ERBB network leads to receptor autophosphorylation of C-terminal tyrosines and recruitment to these sites of cytoplasmic signal transducers that regulate cellular processes such as proliferation, differentiation, motility, adhesion, protection from apoptosis, and malignant transformation [8]. Studies of HER2-overexpressing breast cancer cell lines and human tumors have shown constitutive HER2 phosphorylation and activation [13, 14]. Induced overexpression of HER2 is associated with mammary epithelial cell transformation [15, 16]. These studies indicate that HER2 is a potent oncogene in the mammary gland and a causative factor for breast cancer.

### HER2-targeted Therapies

The humanized antibody trastuzumab and the ATP-mimetic tyrosine kinase inhibitor (TKI) lapatinib are FDA-approved anti-HER2 agents for the treatment of HER2-overexpressing (HER2<sup>+</sup>) breast cancers. As the first approved therapy for treating HER2<sup>+</sup> breast cancers [17, 18], a large amount of clinical data on patient responses to trastuzumab has been obtained. Trastuzumab has been shown to induce tumor regression in 12~35% of heavily pretreated metastatic breast cancers with HER2 overexpression [19–21]. Nevertheless, most metastatic breast tumors with HER2 gene amplification and/or high levels of HER2 protein do not respond to trastuzumab; further, the majority of those cancer that initially respond

eventually relapse, suggesting de novo and acquired mechanisms of therapeutic resistance. The mechanisms of resistance to trastuzumab are not fully understood. However, recent reports suggest that overexpression of the IGF-I receptor [22] or activated EGFR [23] as well as aberrant PI3K/AKT signaling [24] or PTEN deficiency [25] may all result in resistance to trastuzumab. Accumulating evidence suggests that combinations of agents targeted to the HER2 network or other pathways synergizing with HER2 may be beneficial for efficient treatment of HER2<sup>+</sup> breast cancers (reviewed in [26]).

## A Synergy Between TGF $\beta$ and HER2 in Mammary Tumor Progression

### TGF $\beta$ Facilitates Metastasis of Neu-mediated Mammary Tumors

Synergy between TGF $\beta$  and HER2/ERBB2 (neu) was initially demonstrated by crossbreeding mice expressing the Neu oncogene in the mammary gland driven by the mouse mammary tumor virus (MMTV) promoter with either MMTV/ALK5<sup>T204D</sup> mice (expressing a constitutively active mutant of the type I TGF $\beta$  receptor or T $\beta$ RI) [27, 28] or MMTV/TGF $\beta$ 1<sup>S223/225</sup> mice (expressing a constitutively active mutant of TGF $\beta$ 1) [28, 29]. In both bi-transgenic models, overexpression of activated receptor or TGF $\beta$  ligand in the mammary gland of mice also expressing neu accelerates metastases from Neu-induced mammary tumors [28–30]. The Neu/ALK5<sup>T204D</sup> and Neu/TGF $\beta$ 1<sup>S223/225</sup> bigenic tumors exhibit less apoptosis and are more locally invasive and of higher histological grade compared to the neu tumors [27, 29]. The neu/TGF $\beta$ 1<sup>S223/225</sup> mice also appear to have more circulating tumor cells than Neu mice. At the molecular level, higher levels of phosphorylated AKT and mitogen-activated protein kinase (MAPK) are observed in tumors expressing both neu and ALK5<sup>T204D</sup> or TGF $\beta$ 1<sup>S223/225</sup> when compared to tumors expressing neu alone [27, 29].

Loss-of-function experiments have also supported the prooncogenic synergy between TGF $\beta$  and Neu signaling. For example, mice expressing soluble T $\beta$ RII exhibit high levels of this TGF $\beta$  antagonist in circulation, leading to suppression of metastases from neu-induced mammary tumors [28, 31]. Collectively, these data suggest that TGF $\beta$  can accelerate the metastasis of neu-driven mammary tumors, possibly through the synergistic activation of PI3K/AKT and Ras/MAPK pathways with neu-dependent signaling. Moreover, the findings show that neu requires TGF $\beta$  signaling to maximally drive the metastatic progression of mammary tumors.

### Enhanced In Vitro Migration and Invasion Mediated Through HER2 and TGF $\beta$ Synergy

The functional synergy between TGF $\beta$  and HER2 has been characterized using the MCF10A non-transformed human mammary epithelial cell (HMEC) model. A genetic modifier screen in MCF10A cells stably overexpressing transfected HER2 showed that TGF $\beta$ 1 and TGF $\beta$ 3 cDNAs cooperate with HER2 in inducing cell motility and invasion in both 2D and 3D basement membrane cultures [32]. This cooperation between HER2 and TGF $\beta$  correlates with sustained activation of AKT, ERK and p38 MAPK, and is abolished by pharmacological inhibition of PI3K, ERK, or p38 MAPK, as well as by trastuzumab or an integrin  $\beta$ 1 blocking antibody [32, 33]. Taken together, these in vivo and in vitro data suggest that overexpression of HER2 is permissive for TGF $\beta$ -induced signals associated with tumor cell motility and, potentially, metastatic progression.

## Molecular Mechanisms of the Crosstalk between TGF $\beta$ and HER2

Recent mechanistic studies reveal that the synergistic effect of TGF $\beta$  and HER2 on tumor progression is likely a combined result of loss of TGF $\beta$ 's tumor suppressive function and gain of pro-survival and pro-migratory functions through HER2-dependent mechanisms. In these studies, the crosstalk between TGF $\beta$  and HER2 occurs at various levels, including (1)

suppression of Smad-dependent transcriptional regulation, and (2) Smad-independent induction of ERBB ligands. A previous study indicates that the majority of human breast tumors exhibit intact Smad signaling indicated by the phosphorylation of Smad2, whereas lack of Smad2 phosphorylation is associated with poor patient outcome [34]. Thus, both Smad-dependent and -independent mechanisms may be employed by TGF $\beta$  to promote HER2<sup>+</sup> breast cancer.

### Silencing of TGF $\beta$ 's Tumor Suppressive Effect

Smad transcription factors bind DNA with low affinity. Therefore, the contextual function of Smads relies on interactions with other transcriptional factors with more potent and specific DNA binding capacity. We have previously reported that TGF $\beta$  activates the promoter of the tumor suppressor *maspin* by inducing binding of Smads and p53 to an overlapping Smad binding element (SBE)-p53 site [35]. In fact, a number of genes are synergistically regulated by Smads and p53, and many of them are tumor suppressors. A common feature of the promoter regions of these genes is the presence of overlapping or adjacent p53 binding sites and SBEs, usually within a 100-bp segment [36]. This pattern is also observed in the promoter of the mutS homolog 2 (*MSH2*), a tumor suppressor and central component of the DNA mismatch repair (MMR) system [37]. Because efficient regulation of these genes by TGF $\beta$  and Smads requires intact p53 signaling, this TGF $\beta$ -mediated function is abolished in the context of loss of p53, a frequent alteration in multiple neoplasias, including breast cancer. Overexpression of HER2 has been reported to decrease p53 levels via activation of the PI3K pathway and induction of the nuclear translocation of MDM2, an E3 ubiquitin ligase targeting p53 [38]. Our study also indicates that HER2 overexpression impairs p53-dependent transcriptional regulation of MSH2 by TGF $\beta$ , which could be restored by nutlin-3, a small molecule that induces p53 stabilization by inhibiting MDM2-dependent degradation of p53 [37, 39]. Therefore, HER2 may alter Smad-dependent transcriptional regulation through modulating the functional status of p53 in transformed cells. In addition, overexpression of HER2 increases the level of miR-21, a TGF $\beta$ -inducible miRNA that targets and down-regulates MSH2 transcripts [37]. As a result, in HER2-transformed cells, TGF $\beta$  fails to activate the MSH2 promoter but instead decreases MSH2 expression through induction of miR-21 [37]. This downregulation of MSH2 by TGF $\beta$  also contributes to resistance to DNA-damaging chemotherapy agents in cancer cells, as MSH2 is required for the recognition of drug-induced DNA damage, which triggers apoptosis [37].

A recent study by Arnal-Estape et al. indicates that antagonism between two isoforms of the transcription factor C/EBP $\beta$  also alters Smad function in the context of HER2 overexpression [40]. In non-transformed epithelial cells, TGF $\beta$  induces formation of the C/EBP $\beta$ -Smad2/3-E2F4/5 transcriptional regulatory complex on the MYC promoter and inhibits gene transcription [41]. This tumor suppressive mechanism is abolished in HER2-overexpressing breast cancer cells. In this case, HER2 signaling increases the production of C/EBP $\beta$  isoform LIP, which antagonizes the TGF $\beta$ -induced assembly of transcriptional repressor complexes containing Smad and the active C/EBP $\beta$  isoform LAP on the MYC promoter [40]. Thus, the HER2-mediated functional switch between the two C/EBP $\beta$  isoforms, as well as its modulation of p53 activity, may both contribute to the silencing of TGF $\beta$ 's tumor suppressive function, which is largely mediated by Smad-dependent transcriptional regulation (Fig. 1).

In addition to the Smad cofactors p53 and C/EBP $\beta$ , HER2 has also been reported in breast cancer cells to collaborate with ETS transcription factor ER81 to activate the transcription of Smad7, an inhibitory Smad that suppresses Smad2/3-mediated gene regulation [42]. These events may also lead to a general suppression of the anti-proliferative Smad-dependent TGF $\beta$  action in HER2<sup>+</sup> breast cancer cells.

## Potential of HER2-mediated Oncogenic Signaling

Using the HER2-overexpressing MCF10A cell model (MCF10A/HER2), we have shown that addition of exogenous TGF $\beta$  or expression of constitutively active T $\beta$ RI (ALK5<sup>T204D</sup>) induces motility of MCF10A/HER2 cells but not control MCF10A/vec cells [33]. This effect is mediated at least in part by activation of PI3K and involves translocation of HER2 to cell membrane protrusions, where it co-localizes with Vav2, Rac1, Pak1 and the actin cytoskeleton, resulting in prolonged Rac1 activation and enhanced cell survival and invasiveness [43]. By anchoring HER2 to actin fibers, TGF $\beta$  also induces clustering of HER2 with integrin  $\alpha$ 6,  $\beta$ 1 and  $\beta$ 4; this clustering is mediated by focal adhesion kinase (FAK) and is required for TGF $\beta$ -induced motility and oncogenic signaling of HER2 in breast cancer cells (Fig. 1) [44]. We further investigated the mechanism through which TGF $\beta$  activates PI3K in HER2-overexpressing cells, and found that treatment with TGF $\beta$  or expression of ALK5<sup>TD</sup> induces phosphorylation of the TACE/ADAM17 sheddase and its translocation to cell surface, resulting in increased secretion of TGF- $\alpha$ , amphiregulin, and heregulin. In turn, these ERBB ligands enhance HER2-mediated signaling, such as the association of PI3K p85 subunit with the HER2:ERBB3 heterodimers, leading to sustained activation of the PI3K/ AKT signaling pathway (Fig. 1) [45]. Notably, activation of TGF $\beta$  signaling in HER2-overexpressing breast cancer cells also reduces their sensitivity to trastuzumab, likely as a result of PI3K hyperactivation [45].

Another study by Northey et al. indicates that signaling through phosphorylated tyrosine residues 1226/1227 and 1253 of HER2/neu, the sites mediating binding of the ShcA adaptor protein, is essential for the synergistic effect of TGF $\beta$  on the motility and invasion of HER2/neu-over-expressing cells [46]. Suppression of ShcA function using a dominant-negative mutant abrogates the TGF $\beta$ -mediated effect in breast cancer cells [46] suggesting that, similar to PI3K, ShcA is another key adaptor that mediates the crosstalk between TGF $\beta$  and HER2. Although it is not yet clear if TGF $\beta$ -mediated TACE activation and ERBB ligand shedding are the upstream events that cause HER2/neu phosphorylation at these ShcA-interacting tyrosines, this study also supports the model that TGF $\beta$  exerts the proinvasive, and potentially, pro-metastatic effect by modulating the amplitude of HER2/neu-dependent signaling.

In addition to TGF $\beta$ -induced shedding of ERBB ligands, changes in the tumor microenvironment occur by way of HER2-induced expression and secretion of TGF $\beta$ 1 and TGF $\beta$ 3 through a mechanism involving Rac1 activation and JNK-AP1-dependent transcription [47]. Moreover, vascular endothelial growth factor (VEGF), a target of the TGF $\beta$ -Smad transcriptional regulation, is synergistically induced by HER2 and TGF $\beta$  [47]. Thus, the crosstalk between HER2 and TGF $\beta$  not only alters intracellular signaling in cancer cells but also influences other components of the tumor microenvironment through the induction of several pro-invasive growth factors. Targeting these extracellular factors may provide novel therapeutic strategies directed at both cancer-driving oncogenes and the modified tumor microenvironment.

## Clinical Implications of HER2-dependent TGF $\beta$ Action

To understand the clinical relevance of the HER2:TGF $\beta$  crosstalk, we mapped an ALK5<sup>T204D</sup>-induced gene expression signature to an array dataset published by van de Vijver et al. [48] and Chang et al. [49]. The signature correlated with biological and clinical differences among 295 primary breast tumors. Tumors whose gene expression correlated with the active T $\beta$ RI (ALK5<sup>TD</sup>) signature are mostly HER2-positive or basal-like with some luminal B tumors, while the tumors with a negative correlation are predominantly luminal A and normal-like tumors (Fig. 2a) [45]. Patients with tumors expressing the ALK5<sup>TD</sup> signature exhibited a shorter survival than those with tumors that do not express the

ALK5<sup>TD</sup> signature (Fig. 2b). We further explored possible correlations of the ALK5<sup>TD</sup> signature with resistance to trastuzumab by mapping this gene expression signature to an array data set reported by Harris et al. [50] that had been obtained from 22 patients with HER2-overexpressing breast cancer treated with neoadjuvant trastuzumab and vinorelbine. Hierarchical clustering analysis shows that all 3 patients achieving pathological complete response do not share a similar expression pattern with the TGF $\beta$  signature (Fig. 2c & d) [45]. These findings, therefore, support an association between TGF $\beta$  signaling and clinical resistance to trastuzumab. A causal association will require confirmation in clinical studies using combinations of HER2 and TGF $\beta$  antagonists.

As indicated by the studies reviewed herein, the cellular phenotypes induced by TGF $\beta$  are context-dependent and largely edited by the overexpression of HER2, the major pathogenic oncogene in a significant cohort of breast cancers. In HER2-transformed cells, TGF $\beta$  further stimulates HER2 signaling to promote malignancy and may induce resistance to anti-HER2 therapy. Taken together, the documented evidence suggests that simultaneous blockade of the HER2:TGF $\beta$  axis may significantly enhance the efficiency of conventional therapies in patients with HER2-dependent breast cancers.

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## Abbreviations

<b>EGFR</b>	Epidermal growth factor receptor
<b>ERBB</b>	Erythroblastic leukemia viral oncogene homolog
<b>ERK</b>	Extracellular signal-regulated kinase
<b>FAK</b>	Focal adhesion kinase
<b>HER2</b>	Human epidermal growth factor receptor 2
<b>JNK</b>	C-Jun NH <sub>2</sub> -terminal kinase
<b>MAPK</b>	Mitogen-activated protein kinase
<b>MMR</b>	DNA mismatch repair
<b>MSH2</b>	mutS homolog 2
<b>PI3K</b>	Phosphatidylinositol-3 kinase
<b>RTK</b>	Receptor tyrosine kinase
<b>SBE</b>	Smad binding element
<b>TGF<math>\beta</math></b>	Transforming growth factor $\beta$
<b>VEGF</b>	Vascular endothelial growth factor

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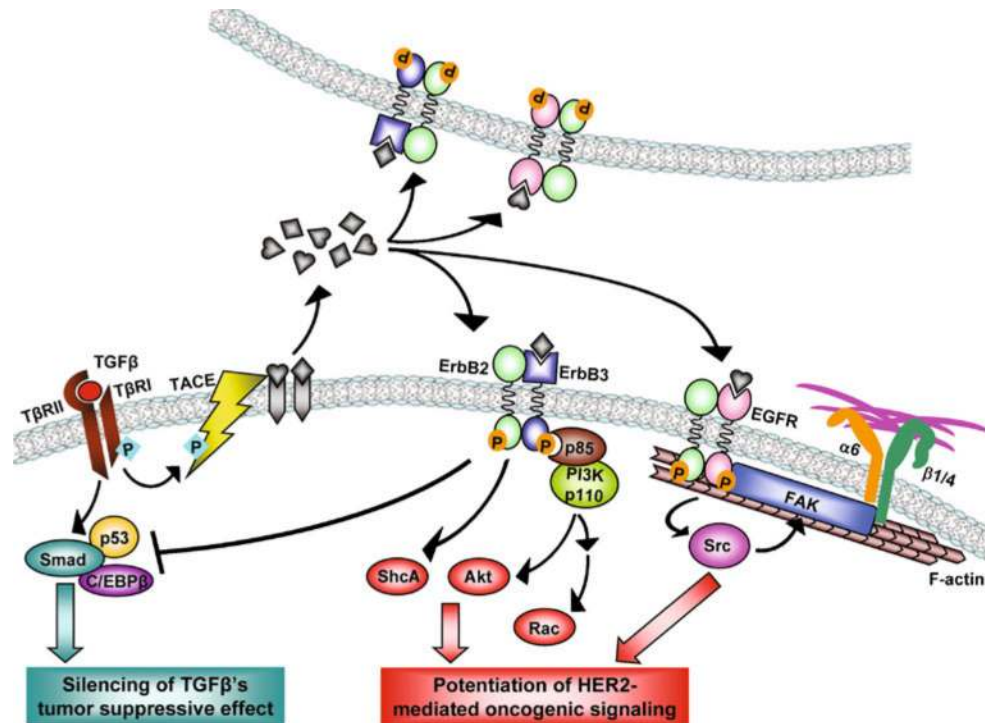
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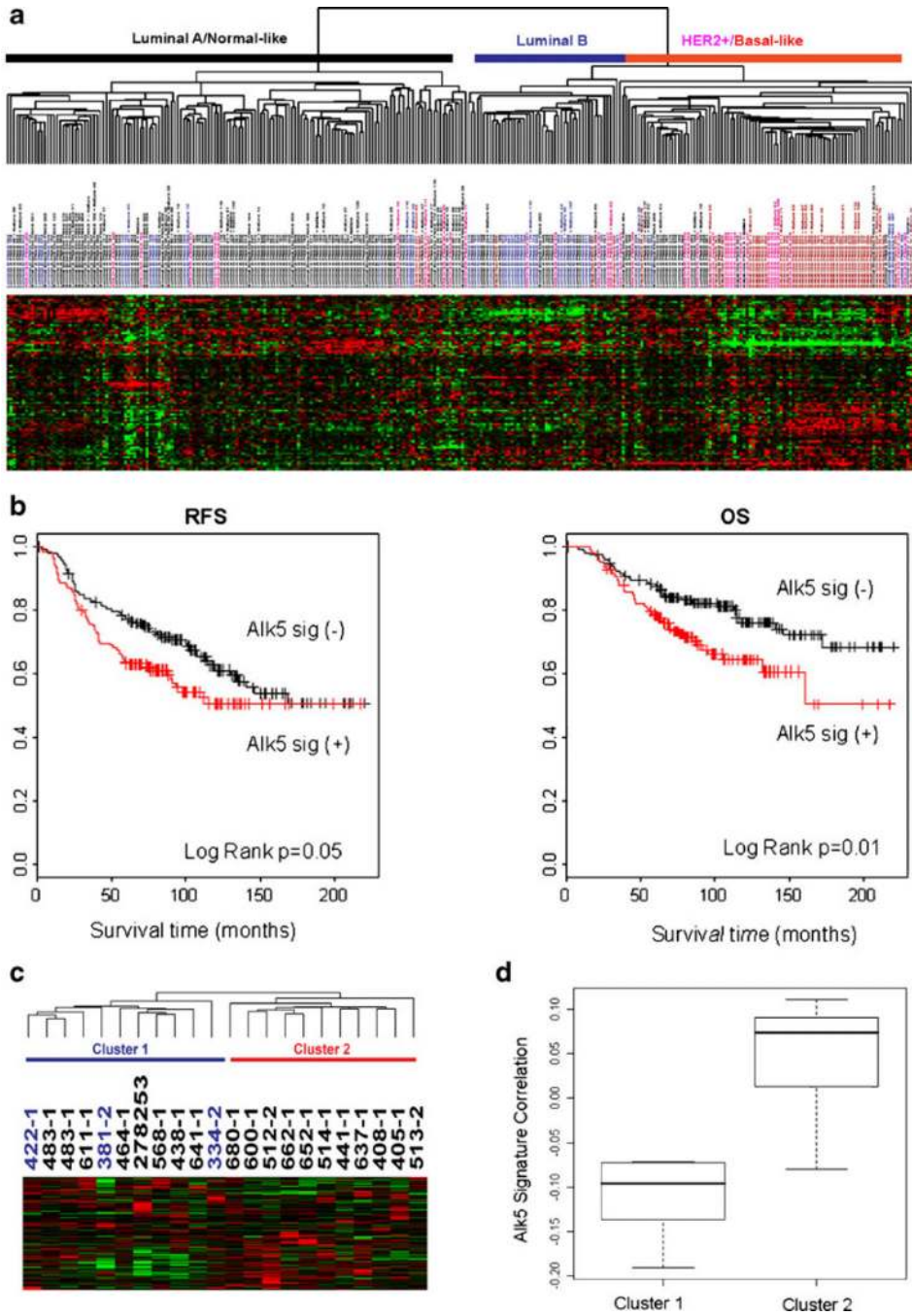
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**Figure 1.** Mechanisms of the pro-malignant crosstalk between TGF $\beta$  and HER2 in breast cancer (based on findings in [35, 37, 40, 43–46]). Signals from extracellular TGF $\beta$  are transduced into cells that express wild-type TGF $\beta$  receptors. Activated T $\beta$ RI induces a set of Smad-dependent cytosolic gene responses, which are impaired by HER2-mediated alterations of p53 and C/EBP $\beta$  activities, as described in [35, 37, 40]. On the other hand, T $\beta$ RI phosphorylates TACE, resulting in its translocation to the cell surface where it cleaves ERBB pro ligands [45]. ERBB ligands will initiate autocrine and paracrine ERBB signaling in adjacent cells. Ligand-induced ERBB signaling, especially in breast cancer cells overexpressing the ERBB signal amplifier HER2, can subsequently lead to activation of the ShcA, PI3K/Akt, and Src/FAK/integrin signaling cascades to promote cancer progression [43–46]. Thus, the crosstalk between TGF $\beta$  and HER2 is the functional sum of silenced TGF $\beta$ 's tumor suppressive effect and enhanced HER2-mediated oncogenic signaling



**Figure 2.** ALK5<sup>TD</sup> signature is associated with clinical outcome in women with breast cancer (figure adapted from [45]). **a** Hierarchical clustering of 295 breast tumors [48, 49] using 90 overlapping genes with the 271-gene ALK5<sup>TD</sup> signature. **b** Kaplan Meier plots for recurrence-free survival (RFS) and overall survival (OS) comparing the two groups of tumors with and without a correlation with the ALK5<sup>TD</sup> signature. **c** Hierarchical clustering of 22 breast tumors from patients who were treated with navelbine and trastuzumab [50] using 190 overlapping genes with the 271-gene ALK5<sup>TD</sup> signature. Cluster 2 shows a

positive correlation with the ALK5<sup>TD</sup> signature. **d** Box-and-Whisker plot of Standard Pearson Correlation between the ALK5<sup>TD</sup> signature and Clusters determined in **c**