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Whole blood transcriptomic profiling identifies molecular pathways related to cardiovascular mortality in heart failure

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Abstract

Aims: Chronic Heart Failure (CHF) is a systemic syndrome with a poor prognosis and a need for novel therapies. We investigated whether whole-blood transcriptomic profiling can provide new mechanistic insights into cardiovascular (CV) mortality in CHF.

Methods and Results: Transcriptome profiles were generated at baseline from 944 CHF patients from the BIOSTAT-CHF Study - of whom 626 survived and 318 died from a CV cause during a follow-up of 21 months. Multivariable analysis, including adjustment for cell count, identified 1,153 genes (6.5%) that were differentially expressed between those that survived or died and strongly related to a validated clinical risk score for adverse prognosis. The differentially expressed genes mainly belonged to 5 non-redundant pathways: Adaptive immune response, proteasome-mediated ubiquitin-dependent protein catabolic process, T-cell co-stimulation, positive regulation of T-cell proliferation and erythrocyte development. These five pathways were selectively related (RV coefficients >0.20) with seven circulating protein biomarkers of CV mortality (FGF23, sST2, adrenomedullin, hepcidin, pentraxin-3, WFDC2 and IL-6) revealing an intricate relationship between immune and iron homeostasis. The pattern of survival-associated gene expression matched with 29 perturbagen-induced transcriptome signatures in the iLINCS drug-repurposing database, identifying drugs, approved for other clinical indications, that were able to reverse *in vitro* the molecular changes associated with adverse prognosis in CHF.

Conclusion. Systematic modeling of the whole blood protein-coding transcriptome defined molecular pathways that provide a link between clinical risk factors and adverse cardiovascular prognosis in CHF, identifying both established and new potential therapeutic targets.

Keywords: Chronic Heart Failure, RNA, T-Cells, Interleukins, Fibroblast growth factor 23, Iron, drug-repurposing

Introduction

The prognosis of patients with chronic heart failure (CHF) remains poor, despite substantial improvements in CHF diagnosis and treatment. Several prognostic risk scores have been developed for CHF[1,2] yet the precise mechanistic links between such clinical risk scores and survival, remain unclear. Furthermore, beyond natriuretic peptides[3], several circulating protein biomarkers that have been shown to be independently associated with prognosis in CHF[4–6] remain to have the underpinning molecular relationships elucidated.

Whole blood transcriptomic profiling is a practical and powerful methodology to study genome-wide biology in large numbers of patients. Despite CHF being a systemic syndrome, no study has investigated whether whole-blood transcriptomic profiles can inform about the molecular processes underpinning the observed relationships between clinical or protein biomarkers and cardiovascular mortality in these patients. If the whole blood transcriptome can be shown to reflect important drivers of mortality, then modelling whole blood global gene expression may help identify clinically relevant pathways to target therapeutics development.[7]

In this study, we report the first large-scale whole-blood transcriptomic profiles from patients with CHF. Our main aims were (i) to contrast such profiles between patients who survived or died, (ii) examine whether this provides new insights into molecular mechanisms through which clinical and protein biomarkers associated with adverse outcome might affect prognosis and (iii) explore the utility of blood based transcriptomic signatures as a novel tool to aid *in vitro* drug-repurposing analysis in CHF.

Methods

Participants. This was a nested case-control study undertaken within the BIOSTAT-CHF study[8]. Briefly, BIOSTAT-CHF enrolled an index cohort of 2,516 patients from 69 hospital centers in 11 European countries between 2010 and 2014. Patients were aged >18y with symptoms of new-onset or worsening CHF, confirmed either by a left ventricular ejection fraction (LVEF) of ≤40% or B type natriuretic peptide (BNP) level >400 pg/mL and/or N-terminal pro B-type natriuretic peptide (NT-proBNP) plasma levels >2000 pg/mL, treated with either oral or intravenous furosemide ≥40 mg/day or equivalent at the time of inclusion. After enrolment and obtaining a blood sample, all participants had their heart failure medication optimized to guideline recommended or maximally tolerated doses. The median follow-up of the cohort was 21 months, with an interquartile range of 15 months. The study complied with the 2008 Declaration of Helsinki and was approved by the relevant ethics committee in each center. All participants gave written, informed consent to participate.

For the current transcriptome analysis, participants were categorized into two groups: **survivors** (those participants who survived during the follow-up period without hospitalization), and **non-survivors** (those participants who died from a CV cause). Cause of death (CV or non-cardiovascular) was adjudicated by a senior clinician (A.A.V.) based on reports of mortality events using previously reported criteria[9]. There were 441 CV deaths (out of a total of 657 deaths), and 1,437 participants survived without hospitalization. After the exclusion of participants with no usable RNA samples, 332 who died and 1061 who survived, were available to study. The two groups, those that survived or died, were

matched for age and sex with an approximate ratio of 1:2 to match the number of arrays available (**Supplementary Figure 1)**.

Generation of transcriptomic profiles. Whole blood was collected into PAXgene tubes (Qiagen) and RNA was isolated using TRizol® (Life Technologies), dissolved in 20 µl RNAse-free water and profiled using the Affymetrix Human Transcriptomic Array 2.0 (HTA, Thermo Fisher Scientific) which contains ~7 million probes to provide a comprehensive view of the transcriptome. Samples were processed following the manufacturer's protocol (Sutton Bonington Array facility, University of Nottingham). Briefly, 250ng of RNA was processed to single-stranded sense fragmented DNA using the WT PLUS Reagent Kit. Five µg of fragmented, end–labelled sense stranded target cDNA was hybridized to each array, and scanned in batches of 24, using a 30007G scanner. The running order used a random list generator to ensure even distribution of cases and controls during profiling. Full details of the processing of the arrays and quality checks of the gene expression data are provided in the **Supplementary Materials.** A total of 944 samples were successfully profiled, of whom 626 (66%) were survivors and 318 (34%) died of a cardiovascular cause.

Statistical and bioinformatics analysis. RNA expression values were derived for 17,748 protein-coding genes (ENSG), then normalized and logarithm (base 2) transformed. We applied a multivariable linear model analysis with group (survival and non-survival), age, sex and estimates of major white cell populations as covariates using the limma package in R, to identify differentially expressed ENSGs. Estimated white cell proportions were derived by applying ABIS[10], with absolute values being scaled to 100 prior to further analysis (see **Supplementary Materials**). P-values were adjusted using the Benjamini-Hochberg False

Discovery Rate (FDR) method; an FDR value of less than 5% was used. Gene ontology (GO) profiles (Biological Processes) were assessed using Metascape[11], with the detectable transcriptome used as the GO background to calculate enriched pathway statistics[12]. The main drivers to GO enrichment were identified by removing associated processes, using the topGO hierarchical analysis package in R.

To assess whether there was a relationship between the observed transcriptomic differences between survivors and non-survivors, and clinical and biochemical risk factors associated with CHF prognosis, we used a reproducible prognostic risk score for mortality developed in BIOSTAT-CHF (BIOSTAT-CHF risk score, BRS).[2] The BRS model considered 42 clinical and biochemical variables, identifying 5 variables (more advanced age, higher blood urea nitrogen and N-terminal pro-B-type natriuretic peptide, lower haemoglobin, and failure to prescribe a beta-blocker) as the strongest independent predictor of mortality with a combined C-statistic of 0.73.[2] We undertook linear correlation analysis of gene expression with the BRS and compared the overlap of these set of genes with those differentially expressed between survivors and non-survivors. The effect of adjusting for the BRS on the latter set of genes was also investigated.

To gain insight into potential protein drivers of the transcriptional changes, we correlated the transcriptomic changes with plasma levels of >150 circulating proteins available in the BIOSTAT-CHF cohort, some of which have been reported to be associated with prognosis in the cohort[5,6]. The majority of proteins were measured using two commercial platforms (Olink Proseek Multiplex and Alere Luminex Panel), with a minority derived from routinely available hospital clinical assays or bespoke assays. Specifically, FGF23 data were generated

using the c-terminal ELISA (Immutopics, Inc., San Clemente, CA, USA) while Hepcidin was measured using the competitive ELISA developed by Kroot *et al*[13]. To examine the relationship between each of the non-redundant gene ontologies, and the protein biomarkers, we estimated the RV coefficient, a multivariate generalization of the squared Pearson correlation coefficient. Pearson correlation coefficients, adjusting for multiple testing (Bonferroni correction), were also calculated for the significant associated proteins and individual genes within the top-ranked ontology categories. For proteins most associated (RV coefficient >0.20) with at least one significant gene ontology, we examined the relationship with CV mortality, using a Cox proportional hazards model.

Finally, to explore the potential of our approach to identify new drug treatments, we utilized the CMap-L1000v1[7] *in vitro* drug signature database (https://clue.io/) as previously described[14]. Briefly, to represent the prognosis-related transcriptomics signature we used the 60 most upregulated and 60 most down-regulated genes that were also present in the CMap-L1000v1 database. Choice of 60/60 genes is empirical but informed by our recent work on insulin resistance drug repurposing[14] that yielded a drug list composed of ~50% true-positive drug classes and/or drug targets. The matching score was calculated by aggregating the transcriptional pattern across 9 independent cell lines for >2,500 compounds, many of which include FDA approved drugs.[14] For each matching drug, the known protein targets were identified using PubChem and the small molecule suite[15]. A network of significant pathway interactions was derived using metascape.org, using the CHF survival associated genes and the top ranked known protein target of each drug as input values, and the detectable transcriptome as background.

Results

Subjects

The two groups (survivors and non-survivors) were, by design, closely matched for age and sex (**Table 1 and Supplementary Figure 1**). Patients were predominantly Caucasian; there was no difference in hypertension but there was a higher prevalence of both Type 2 diabetes mellitus (T2DM) and CHF of ischemic aetiology, in the non-survivor group. Baseline use of CHF medication was similar in both groups, except for a greater use of beta-blockers in the survivor group. The echo-estimated left ventricular ejection fraction was similar between the groups (≈31%) as was heart rate. However, patients that died had worse symptoms, lower systolic and diastolic blood pressure and higher plasma concentrations of natriuretic peptides, and lower eGFR, haemoglobin and BMI, at baseline.

Whole blood transcriptomic differences between survivors and non-survivors

Laboratory-measured *total* leucocyte count did not differ between those that survived or died (**Table 1**). However, counts modelled for individual blood cell types using specific gene expression markers (see **Methods**), identified significant reduction in T-cells (-17%, $p<1x10^{-10}$) and B-cells (-11%, $p<1x10^{-3}$) in the non-survivor group, and an increase in neutrophils (+5%, $p<1x10^{-5}$) and basophils (+14%, $p<1x10^{-3}$, **Supplementary Figure 2A)**. After including the estimated cell subtypes as covariates in the analysis together with age and sex, we found that 1,153 genes (6.5%) were differentially expressed (FDR<5% and >5% difference, 557 up-regulated and 596 down-regulated) between survivors and non-survivors (**Figure 1A and Supplementary Table 1**). Up and down-regulated genes often belonged to the same biological pathways (**Supplementary Figure 2B**) and mapped to several interrelated biological pathways (**Figure 1B and Supplemental Table 2**). The processes most responsible

for the GO results were identified by removing the redundant topological features of the GO structure (**Supplementary Figure 3**). This identified five main 'driver' GO categories, each with a robust fold enrichment (FE) and low False Discovery Rate (FDR): Adaptive immune response (FE=2.3, FDR<0.1%), Proteasome-mediated ubiquitin-dependent protein catabolic process (UPP, FE=1.7, FDR<3%), T-cell co-stimulation (FE=3.9, FDR<0.1%), Positive regulation of T-cell proliferation (FE=3.0, FDR<0.1%) and Erythrocyte development (FE=4.3, FDR<0.1%).

The differentially expressed genes in each of these pathways are shown in **Figure 2**. Overall, the pattern of changes in gene expression indicate an impairment of the adaptive immune response, down-regulation of T-cell co-stimulation and disruption of positive regulation of T-cell proliferation in non-survivors. In contrast, the UPP pathway and the erythrocyte development pathway were both activated in non-survivors (see **Discussion**).

BIOSTAT-CHF prognostic risk score and transcriptomic changes

Linear regression analysis of gene expression versus the BRS[2] identified 1,086 correlated genes, and these predominantly belonged to the same pathways as the genes differentially expressed between survivors and non-survivors (**Supplementary Figure 4**). On an individual gene level basis, 390 genes were common to both lists. Inclusion of the BRS as a covariate in the analysis of the expression differences between survivors and non-survivors removed all but one of the gene expression differences between groups. This suggests that the observed variation in the blood transcriptome is strongly related to clinical factors associated with prognosis.

Transcriptomic changes and circulating protein biomarkers

To examine whether circulating proteins represented potential drivers of the transcriptomic changes, and therefore potential intermediaries between the clinical prognostic variables and the observed gene expression differences, we modelled the relationships between >150 circulating proteins, and the five top-ranked GO categories. While most demonstrated limited association (**Supplementary Table 3**), seven proteins demonstrated an RV coefficient of 0.2 or greater, with at least one of the five gene ontologies (**Table 2**). These were, in order of association: Fibroblast Growth Factor 23 (FGF23), soluble ST2 receptor (sST2), Adrenomedullin (ADM), Hepcidin (HEPC), Pentraxin-3 (PTX3), WAP 4-Disulfide core domain 2 (WFDC2) and Interleukin 6 (IL-6). Notably, six of these proteins positively covaried with each other, while HEPC demonstrated a negative correlation, especially with FGF23 (**Supplementary Figure 5**). These seven proteins also demonstrated a significant univariate association with mortality in the present nested case-control sample (**Table 2**), as previously reported for some of these proteins in the full BIOSTAT-CHF cohort[5,6]. Notably, although T2DM was more common in non-survivors at baseline, we did not observe any pathway-level association with HbA1c (**Supplementary Table 3**).

Co-expression analysis

To further delineate the inter-relationships between genes within each of the five GO categories and their relationship with the protein biomarkers, and how these might relate to prognosis, we created Pearson correlation heatmaps for each category, for survivors and non-survivors separately (**Figure 3 and Supplementary Figure 6**). To facilitate identification of altered co-expression relationships between survivors and non-survivors we also created differential co-expression plots (**Supplementary Figures 7 and 8**). The direction of gene co-expression was largely conserved between survivors and non-survivors, partly speaking to

the robustness of the methodologies utilized. However, these analyses highlighted several individual examples of altered gene co-expression, e.g. the iron transporter, TFRC (Transferrin Receptor), was upregulated in non-survivors (**Figure 2**) and correlated to IL6ST (gp130) expression only in non-survivors (**Figure 3B** and **Supplementary Figure 8B**). Gene expression within the UPP pathway was subject to numerous shifts in the correlative structure (**Figure 3 and Supplementary Figure 7B**) with genes like HECTD3 (upregulated in non-survivors) becoming more positively co-expressed with other members of the pathway (except for SMURF2 and RNF216). Likewise, the correlations between the seven protein biomarkers (FGF23, sST2, ADM, HEPC, PTX3, WFDC2 and IL-6) tended to be stronger in the non-survivors, except for IL-6 (**Figure 3A** versus **3B**). The correlation between the seven individual proteins and individual genes varied substantially (**Supplementary Figures 7-8)** and this provided insight into some likely connections between the transcriptome profile and survival (See **Discussion**).

Using mortality related transcript patterns to identify new drug targets

A search, using the combined 60 most up-regulated and 60 most down-regulated genes between survivors and non-survivors that mapped to the iLINCS database, against >2,500 *in vitro* drug signatures found 45 matching compounds (**Supplementary Table S4**). This included 29 that reversed the mortality associated gene expression pattern, and these drugs had 47 known protein targets (<1uM potency). Combined pathway analysis of the drug targets and the top regulated mortality associated genes established that the 47 proteins mapped to biological processes identified by the CHF mortality related transcriptome. This included the 'immune response-activating cell receptor signalling' and 'lymphocyte activation' pathways (**Figure 4**).

Discussion

Whole blood transcriptomic profiling is a potentially powerful methodology to study genome-wide biology directly in patients. Here, in the first prospective study of this type in heart failure, we provide novel insights into molecular processes associated with adverse cardiovascular prognosis. Specifically, we show that such profiles differ at baseline between those patients with CHF who died of a cardiovascular cause (over a median follow-up of 21 months) compared to those that survived, with more than 1 in 20 of the detectable protein-coding transcriptome showing a difference. Although the overall fold differences in gene expression between survivors and non-survivors were small, these differences are likely to be driven by a subset of immune cells and therefore the fold change values would be 'diluted' by other cell types contributing to the global RNA profile. Thus, the magnitude of the measured differences should not be equated with their biological significance. Indeed, the differences could be mapped to specific biological pathways, several of which have been postulated to be disrupted in CHF. Importantly, we show that the differentially regulated transcriptome is strongly related to clinical and protein predictors of survival. While these findings are correlative, they highlight potential mechanisms and intermediaries through which the risk related to clinical factors is mediated. Finally, we provide initial evidence that the mortality-associated RNA pathways in CHF can be reversed *in vitro* by drugs, some already approved for clinical use, identifying them – and their targets – as being potentially relevant to drug development for CHF.

Altered biological pathways and prognosis in heart failure

In cross-sectional analysis, lymphopenia has been associated with more severe NYHA class and poorer outcomes in CHF.[16,17,] Our study provides further detailed evidence that CHF

patients with a poorer prognosis not only have fewer T- and B-cell, but they also have an altered immune cell phenotype. Specifically, T-cell receptor (TCR) signal-transducing molecules such as CD3 epsilon and gamma, as well the co-stimulatory receptor CD28, are down-regulated in non-survivors. Further, a number of genes encoding protein adaptors of TCR signals (such as Lck, Zap70, PKC-θ, ITK and SKAP1), as well as genes that act to amplifying TCR signals such as CD6[18] were all down-regulated (**Figure 2**). Many of these 'adaptative immune response' pathway members, and those involved in TCR signalling in particular, were negatively correlated (**Figure 3**) with both Arginase gene transcripts (ARG1 and ARG2); which were 14-16% more abundant in those that subsequently died (**Supplementary Table 1**). Arginase degrades arginine and decreased arginine, preceded by a measurable increase in arginase activity, is associated with poorer 6-month outcomes in patients with ST-segment elevation myocardial infarction[19]. Arginase gene expression, induced by lactate production consequent to hypoxia, can limit T-cell proliferation[20]. A negative association between ARG2 and the pro-survival IL-7 receptor (IL7R/CD127) was observed in survivors (**Figure 3A**) potentially linking arginine metabolism and IL7R signalling role in T-Cell survival and memory responses[21]. These observations provide support for altered metabolism contributing to the altered T-cell molecular profile, which our transcriptome-wide analysis indicates is a central feature of those patients that died.

Increased net catabolic activity has long been associated with poorer prognosis in heart failure[22] and our transcriptomics approach also identified that upregulation of the 'proteasome-mediated ubiquitin-dependent protein catabolic processes' pathway (UPP) occurs in circulating blood cells (**Figure 2**). Siah E3 ubiquitin protein ligase 2 (SIAH2) was one of the most upregulated genes in the non-survivor group (28%, $p=9x10^{-7}$) and it mediates

degradation of heme oxygenase-1[23], a cardioprotective enzyme involved with immunomodulation[24]. HECTD3 – which has a role in both cardiac hypertrophy[25] and T helper-17 development[26] – was expressed to a greater extent in those that died, showing a stronger (positive) co-expression (**Supplementary Figure 7B**) with other members of the activated UPP pathway (**Figure 2**). In contrast SMURF2 and RNF216 (two down-regulated genes in those that died) was negatively associated with HECTD3 (**Figure 3B**). This is interesting as SMURF2 is a E3 ubiquitin ligase targeting degradation of pro-inflammatory TGFB/SMAD signalling, while RNF216 contributes to ubiquitin mediated protein degradation of TLR4[27]. Presumably targeted acceleration of protein degradation, through UPP, is required to facilitate proinflammatory processes and these more specific events may represent more tractable therapeutic targets.

It is plausible that one driver of the catabolic profile in those that died, was disrupted iron homeostasis. As discussed below, altered iron regulatory protein, HEPC (which regulates ferroportin capacity[28]) and induction of erythrocyte development revealed a number of key observations. For example, there was a strong positive association with the up-regulated transferrin receptor (which imports iron into cells) and up-regulated APFL (aprataxin PNK-like factor) - a ubiquitous DNA damage response enzyme[29] potentially responding to inappropriate iron accumulation[30]. These observations indicate that molecular processes associated with cachexia in CHF, previously identified in cardiac and peripheral tissues[22], are also evident in the whole blood transcriptome where a pervasive role for disrupted iron homeostasis appears across several of the altered pathways.

Do the mortality associated protein biomarkers drive the transcriptional responses to CHF?

As the seven protein biomarkers associated with the transcriptomic changes co-varied with each other (**Supplementary Figure 5**), it is more challenging to identify which proteins are directly linked to the transcriptome differences, particularly as most have plausible mechanistic roles in CHF. For example, PTX3 can skew T-cell differentiation towards the IL-17 producing phenotype[31] while ADM has a positive impact on the endothelial integrity and anti-inflammatory properties[32]. FGF23, a protein secreted that regulates calcium and phosphate homeostasis[33], is produced by osteocytes and cardiac cells, and promotes hypertrophy and fibrosis in a calcineurin-dependent manner[34]. WFDC2 (also known as HE4) suppresses osteopontin and promotes cell survival in an osteopontin- and IFN-γ-dependent manner[35]. WFDC2 is also linked to increased ARG1 activity and immune suppression[36].

Exploration of single transcript and protein biomarker relationships does, however, provide several clues, particularly for the altered T-cell biology and iron homeostasis (**Figure 3 and Supplementary Figure 6**). For example, HEPC links to both altered UPP activity and altered T-cell biology. HEPC demonstrated a strong association with the UPP and 'erythrocyte development' pathways (**Figure 3 and Supplementary Figure 6**). Notably, the association between Hepcidin protein and expression of genes from the UPP pathway was more striking in the non-survivors (**Figure 3B**, e.g. GCLC, PSME4, SIAH2, TBL1XR1, YOD1 and ZER1). In contrast, Mucolipin (MCOLN1) was negatively associated with HEPC and one of the few upregulated genes in the 'adaptive immune response' pathway. MCOLN1 has an important role in calcium mediated cardiac remodelling[37] and is also implicated in iron homeostasis[38]. Together, these examples, and those listed above, imply that alterations in iron status may be a key integrator for the immune related changes in transcriptome and catabolic pathway activation. sST2 was also up-regulated in those that died, consistent with its relationship

with outcome following myocardial infarction where it subdues IL-33 signalling[39]. sST2 levels were positively correlated with the up-regulated erythrocyte development pathway and of note, in the non-survivor group, the increased 'erythropoiesis development' signature was accompanied by reduced haemoglobin ($p<1x10^{-6}$) and iron (**Table 1**). IL-33 can be cardioprotective and is released upon tissue damage, stimulating T-helper type 1 (Th1), Th2 and pro-inflammatory Th17 cells[40], such that loss of IL-33 through increased sST2 may help explain the observed immune related transcriptome profile. Overall, our analysis indicates that each of these seven survival-related proteins could drive the transcriptome response, coupling immunological and iron homeostasis together with clinical risk factors.

Using mortality related transcript patterns to identify new drug treatments

Not all circulating protein biomarkers or regulated genes make relevant therapeutic targets. For instance, despite being modulated in CHF, targeting TNF-α or IL-6 pathways has not led to improved outcomes in heart failure or been found to reduce major cardiovascular events[41]. Notably, the present analysis did not rank either of these cytokines as a dominant feature of the mortality-related pathways. Instead, our analysis indicates that antagonizing FGF23 signalling and/or down regulating sST2 expression (or its binding to IL-33) represent more enticing targets. Beyond these individual proteins, use of transcriptome signatures represents a promising alternative strategy to discover therapeutics and pathways that can modulate diseases[7,14,42]. In fact, many of the protein targets of the 29 drugs that reversed the mortality related transcriptome *in vitro*, are not regulated in patients, but rather represent members of the same pathways regulated in those that died (**Figure 4**). Notably, several positively acting drugs are known to target heart failure related molecular processes (**Supplementary Table 4**), and several have *in vivo* support for their potential utility in CHF,

including a monoamine oxidase inhibitor which can be cardioprotective[43], forskolin which elevates cAMP and cardiac function[44], and the experimental epigenetic drug, chaetocin (a histone lysine methyltransferase inhibitor), which appears cardioprotective[45]. Our findings encourage a further appraisal of the potential benefit in CHF of some of these drugs and pathways.

Limitations

Our study has several limitations. Despite being the largest whole blood genome-wide transcriptomics study in CHF conducted in a very well-characterised cohort, further large scale independent transcriptomic studies are required to validate our observations. An important component of our strategy was to adjust for differences in cell counts between survivors and non-survivors. We modelled changes in individual white cell subtypes using the transcriptomics data and studies using fresh blood samples from patients with CHF are needed to fully explore the changes we predicted. Most patients in BIOSTAT-CHF had heart failure with reduced ejection fraction and whether our findings extend to those with preserved ejection fraction remains to be determined. Finally, our results are correlative and experimental studies are required to confirm the direct involvement of the various pathways we have identified in adverse prognosis in CHF.

In conclusion, our analysis provides evidence that whole blood transcriptomics can identify molecular pathways that associate with CV mortality in CHF. These reflect clinical factors that are associated with poor prognosis and may be driven by specific intermediary protein factors. We demonstrate the potential utility of such transcriptomic profiling for identifying novel therapeutic targets for CHF.

Author contributions

The BIOSTAT-CHF study was conceived and led by SDA, CCL, MM, FZ, GF, DJvV, JGC, LLN, NJS and AAV (PI). The transcriptomic nested case-control study was designed by MN, SPRR, AK and NJS. SPRR undertook the case-control matching. RNA extractions and transcriptomic profiling were conducted by AK, M C-U, MP and STM. Data quality control and normalization were undertaken by MN, CPN and JAT. Data analyses were carried out by MN and JAT. Interpretation of the findings and additional bioinformatics analysis were undertaken by JAT, MN, SH, TRW, FM-B and NJS. The paper was drafted by JAT, MN, FM-B and NJS. All authors reviewed and revised the paper and approved its submission for publication. MN, JAT and NJS are guarantors of the data (GSE181114).

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Conflicts of interest

The authors have no conflicts of interest or disclosures pertaining to this article.

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Table 1: Demographic, clinical and laboratory variables for study groups

Variable	Survivor (N = 626)	Non-Survivor (N = 318)	p-value
Demographics			
Sex (male)	75.7 (474)	74.5 (237)	0.748
Age (years)	71 (10.7)	71.4 (11)	0.584
BMI (kg/m^2)	27.7 (6.2)	26.6 (7.3)	0.206
Clinical profile			
NYHA classification			<0.001
1	2.9 (18)	0.9 (3)	
2	44.2 (277)	22.6 (72)	
3	41.2 (258)	58.5 (186)	
4	8.6 (54)	14.2 (45)	
LVEF (%)	31.6 (9.9)	31.3 (12.4)	0.747
Heart Rate (bpm)	79.2 (20.2)	80.4 (19.4)	0.386
Systolic blood pressure (mmHg)	128.0 (21.7)	121.1 (22.3)	<0.001
Diastolic blood pressure (mmHg)	76.6 (13.7)	71.7 (12.3)	<0.001
HF history			
Ischemic aetiology	61.2 (340)	70.3 (206)	0.01
HF hospitalization in previous year	22.7 (142)	42.8 (136)	<0.001
Medical history			
Hypertension	65.2 (408)	63.2 (201)	0.599
Diabetes Mellitus	28.3 (177)	39.0 (124)	0.001
Medication at baseline			
ACE inhibitors or ARB	74.8 (468)	69.2 (220)	0.081
Beta blockers	84 (526)	77 (245)	0.011
Mineralocorticoid Receptor Antagonist	51.3 (321)	50.9 (162)	0.977
Laboratory measurements			
Haemoglobin (g/dL)	13.4 (1.8)	12.6 (1.8)	<0.001
Erythrocytes (million cells/mcL)	4.5 (0.6)	4.4 (0.9)	0.31

Leucocytes (x109/L)	7.8 [6.6, 9.3]	7.8 [6.4, 9.6]	0.908
eGFR (mL/min/1.73m2; CKD-EPI)	62.0 [46.9, 76.7]	52.3 [34.8, 69.4]	<0.001
BNP (pg/mL)	174.7 [78.3, 371.1]	338.1 [176.2, 652]	<0.001
NT-proBNP (ng/L)	2005 [949, 4642]	4121 [2332, 9275]	<0.001

Continuous variables are expressed as mean (SD) or median [interquartile range]. Categorical variables are expressed as % (N). BMI, body mass index; NYHA; New York Heart Association; LVEF, Left ventricular ejection fraction; ACEi, angiotensin converting enzyme inhibitor; ARB; angiotensin II receptor blocker; CKD-EPI = Chronic Kidney Disease Epidemiology Collaboration equation; eGFR = estimated glomerular filtration rate; BNP, Brain natriuretic peptide; NT-proBNP, N-terminal B-type natriuretic peptide.

Table 2: Top ontologies and RV coefficient estimates with circulating protein biomarkers

GO Pathway (number of genes)	FGF23	sST2	ADM	HEPC	PTX3	WFDC2	IL-6
Adaptive immune response (56)	0.20	0.25	0.20	0.04	0.22	0.21	0.20
T-cell costimulation (14)	0.16	0.23	0.21	0.01	0.22	0.20	0.18
Positive regulation of T-cell proliferation (18)	0.25	0.25	0.22	0.08	0.21	0.19	0.18
Erythrocyte development (11)	0.17	0.09	0.13	0.18	0.04	0.04	0.04
Proteasome-mediated ubiquitin-dependent protein catabolic process (47)	0.30	0.18	0.21	0.22	0.10	0.09	0.09
Univariate association with Survival (HR)	1.64	1.70	1.92	0.84	1.57	1.88	1.43
	$p<1x10^{-34}$	$p<1x10^{-20}$	$p<1x10^{-22}$	$p<1x10^{-4}$	$p<1x10^{-9}$	$p<1x10^{-28}$	$p<1x10^{-12}$

The table presents the RV coefficients - calculated between the expression of all genes identified within each of the top-ranked GO pathways and the levels of plasma protein biomarkers – for those proteins that demonstrated at least a 0.2 or > relationship with one of the 5 top-ranked GO pathways (all $p<0.001$). They are listed in rank order from the RV analysis as follows, FGF23= Fibroblast Growth Factor 23, sST2= soluble ST2 receptor (decoy receptor for IL-33), ADM = Adrenomedullin, HEPC = Hepcidin, PTX3 = Pentraxin-3, WFDC2 = WAP 4-Disulfide core domain 2 and IL-6 = Interleukin 6. The number of genes in each GO pathway is listed in brackets. FE is the fold enrichment over the GO database.

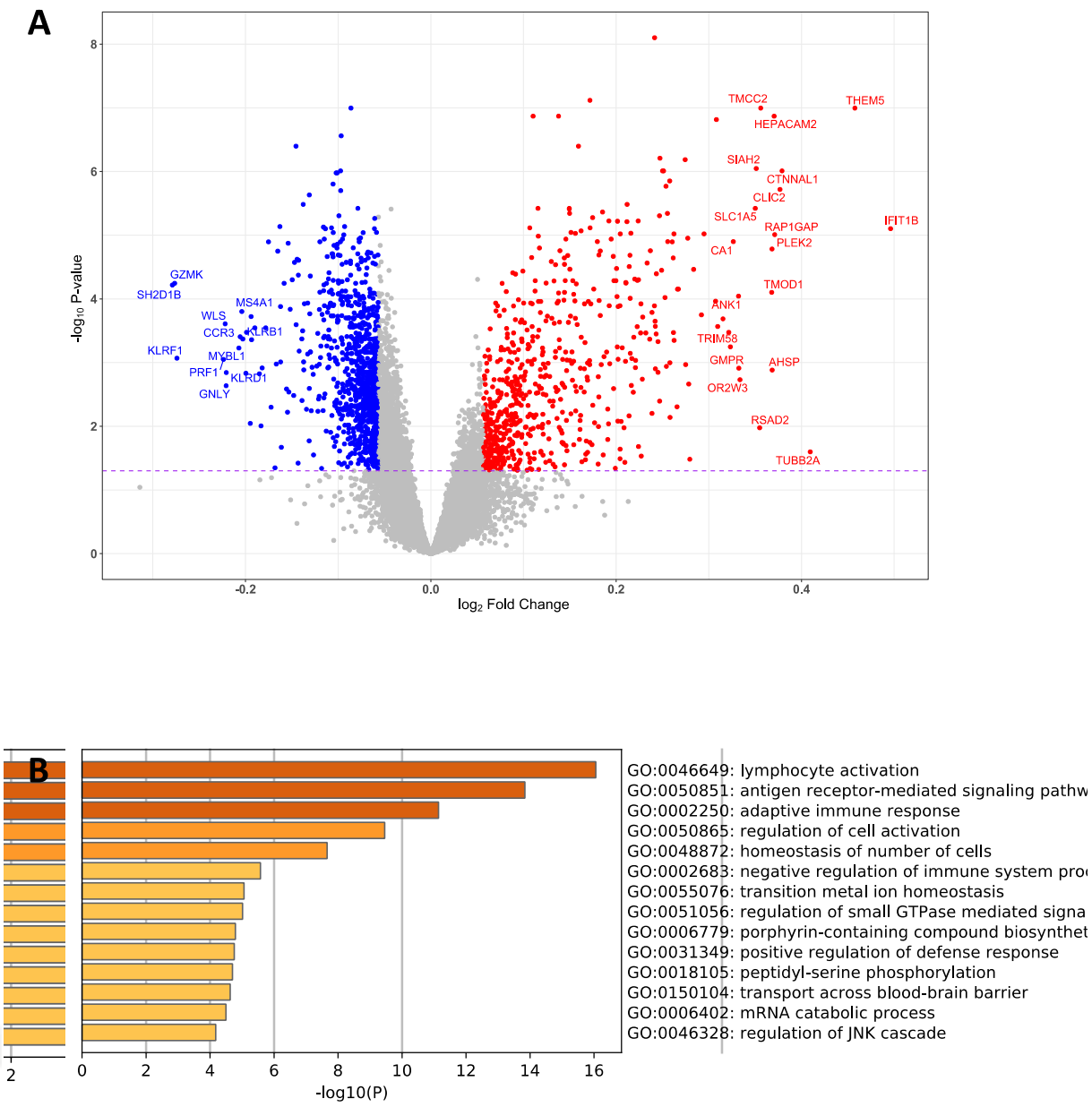

Figure 1. Differential Gene Expression and Biological Process Gene Ontology analysis. A) A volcano plot of the differential gene expression pattern between groups, calculated using limma and the following model; ENSG ~ Age + Sex + log(Sum T-Cells) + log(Sum B-Cells) + log(Neutrophils) + Log(Basophils) +Group. **B)** Global gene ontology (Biological Processes) analysis carried out using Metascape and the list of 1,153 differentially regulated genes. The 17,748 detected protein-coding genes were used as the background and the pre-analysis settings were; threshold >2.0 and non-adjusted p-values threshold p<0.0001. The plot x-axis is p-values (log base 10) while the displayed categories had an FDR=1.4% or better (See Supplementary Table 2).

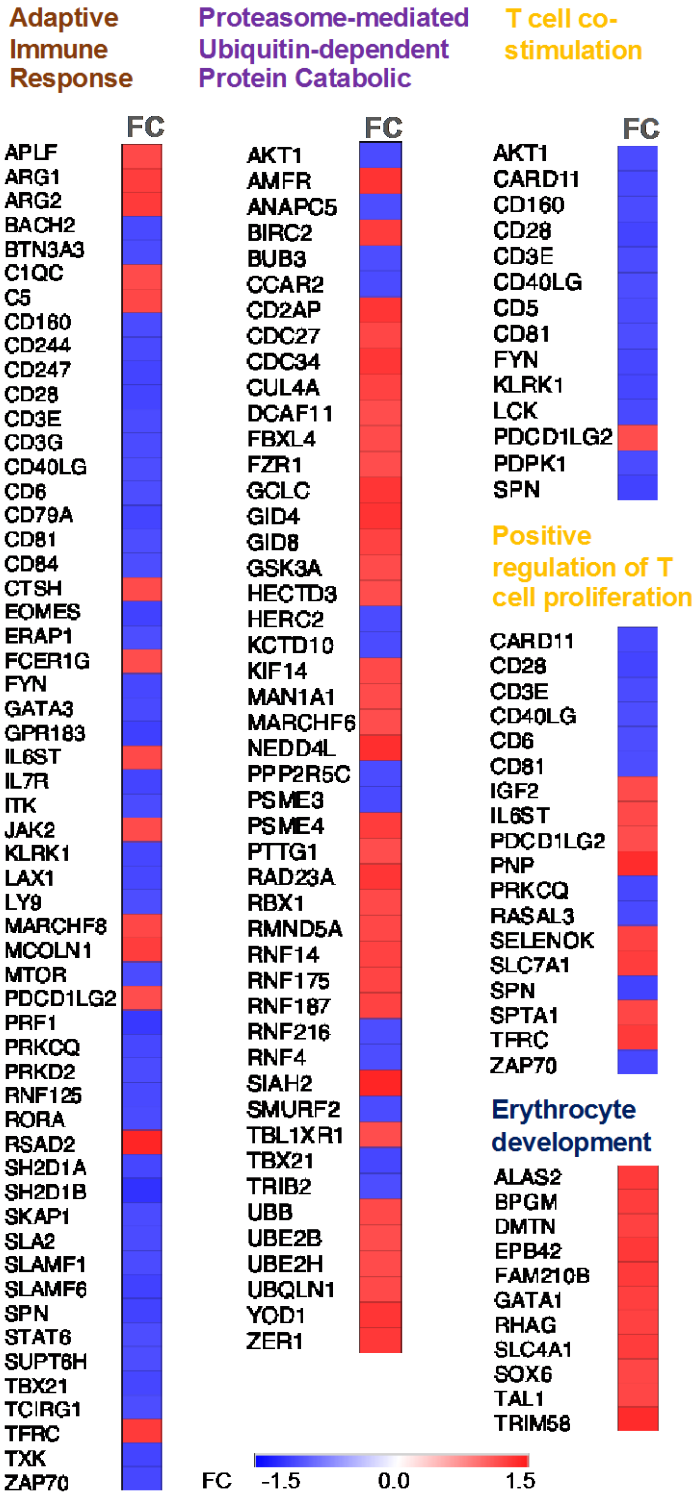

Figure 2. Individual Gene Expression responses within the topology adjusted top-ranked GO categories. Differential gene expression is presented across the 5 topology adjusted gene ontology categories identified using weighted Fisher's exact test, fold enrichment calculations and BH correction of Fisher's test statistics using the R package topGO. FC = fold change.

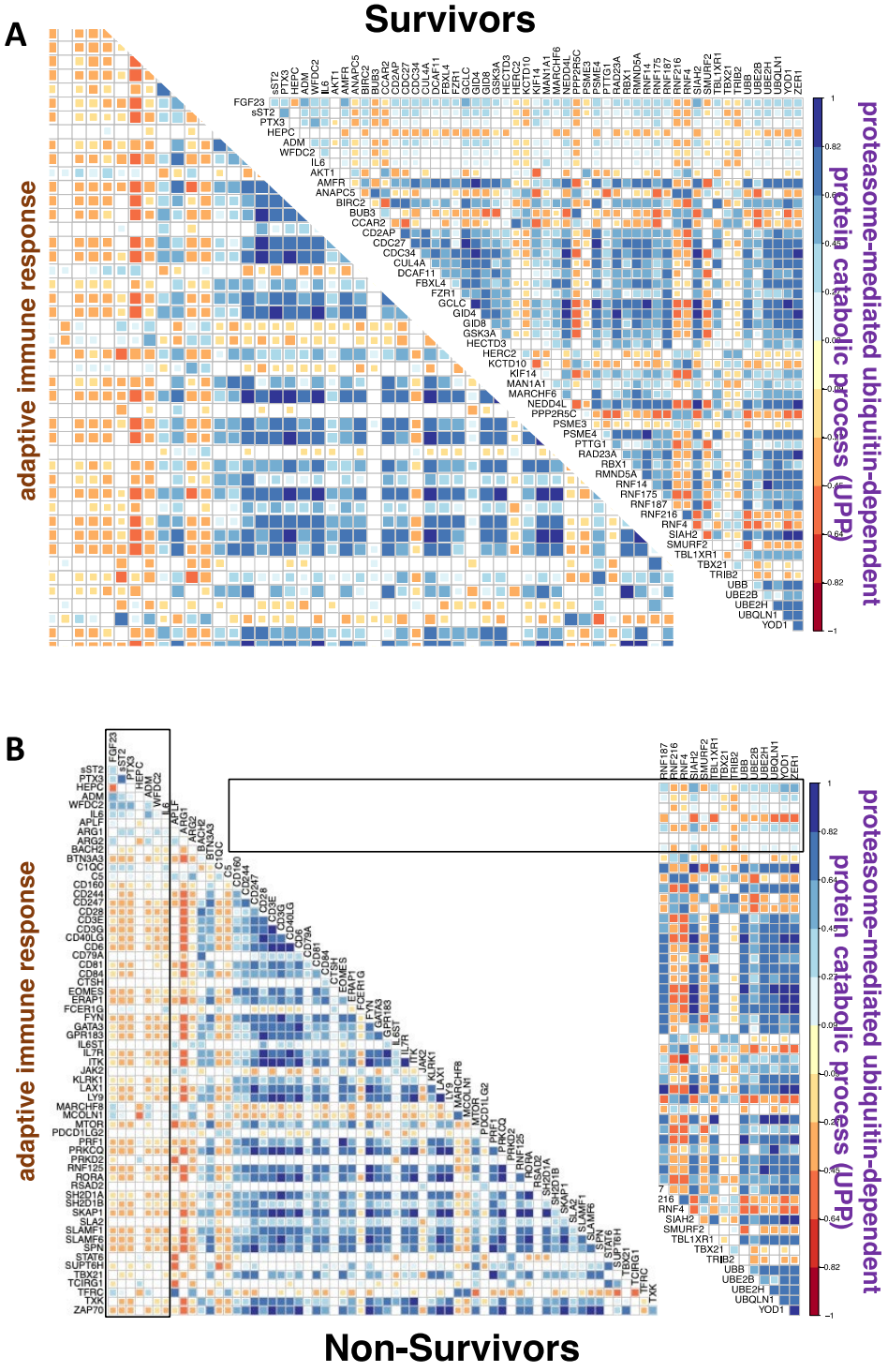

Figure 3. Top GO pathway gene and protein biomarkers inter-relationships. Gene expression was correlated with the top protein biomarkers using Pearson correlation coefficients for the two largest significant pathways 'adaptative immune response' and 'proteasome-mediated ubiquitin-dependent protein catabolic process' – the other three top ranked GO pathways are presented in Supplementary Figure 6. The protein values are enclosed by a black oblong box. Data is plotted separately (grey boxes) for survivors (n=626) and non-survivors (n=318). Correlation values are represented by colour and are plotted for significant correlations (Bonferroni corrected threshold, $p<1.5x10^{-4}$).

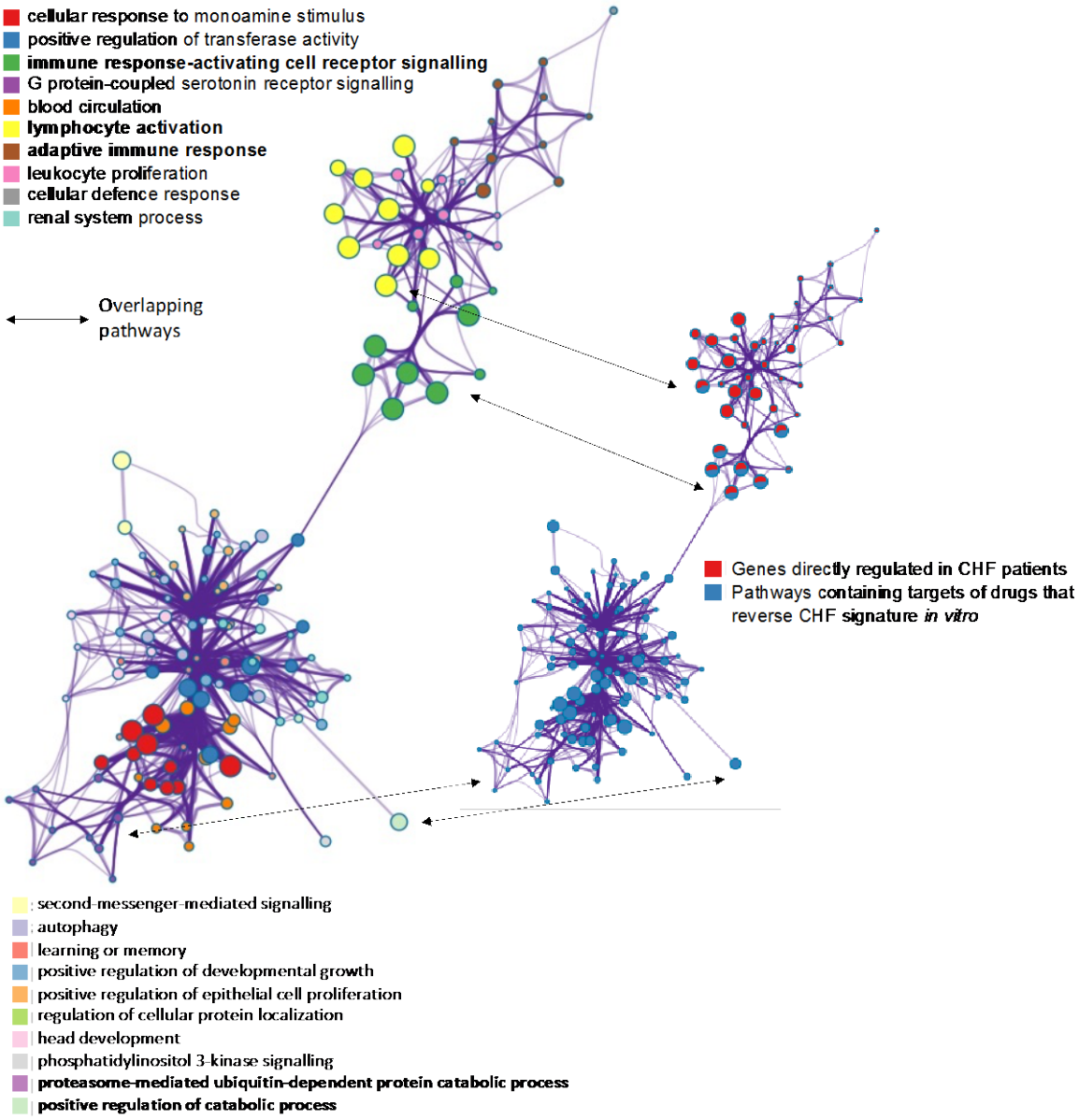

Figure 4: Pathway level overlaps between survival related genes and the protein targets of drugs that reverse the survival related gene expression signature in vitro. A network of significant pathways ($p=10^{-18} – 10^{-4}$) was derived using metascape.org. The input genes were the 120 CHF survival associated genes used to identify drugs that regulate the CHF signature *in vitro* (https://clue.io/) and the 47 protein targets of the 29 drugs (identified using PubChem and the small-molecule-suite) that reverse the CHF signature *in vitro.* There are two identical plots. The large plot, on the **left-hand** side, presents the significant pathways for this combined gene list. Edges represent connected GO biological processes (>0.3), and nodes within each cluster are coloured/named by their most statistically enriched GO term, scaled in size by the total number of terms represented. The smaller plot, on the **right-hand** side, is same network structure but now colour coded by input list membership. This identifies if the drug targets appear within CHF-survival associated pathways (pathways common to Figure 2) or whether the drug target falls within a pathway more indirectly connected with the patient transcriptomic signature. Each node is presented as a pie-chart, with the 'slices' coloured and scaled to indicate which gene list the terms originate from, and what proportion the lists contribute to the ontology groupings.